



## Rate of establishing the gut microbiota in infancy has consequences for future health

Shaillay Dogra<sup>1,†</sup>, Olga Sakwinska<sup>2</sup>, Shu-E Soh<sup>3</sup>, Catherine Ngom-Bru<sup>2</sup>, Wolfram M Brück<sup>2</sup>, Bernard Berger<sup>2</sup>, Harald Brüßow<sup>2</sup>, Neerja Karnani<sup>1</sup>, Yung Seng Lee<sup>1,3</sup>, Fabian Yap<sup>4</sup>, Yap-Seng Chong<sup>1,5</sup>, Keith M Godfrey<sup>6</sup>, and Joanna D Holbrook<sup>1,\*</sup>

<sup>1</sup>Singapore Institute for Clinical Sciences (SICS); Agency for Science and Technology Research (A\*STAR); Singapore; <sup>2</sup>Nestlé Research Center; Lausanne, Switzerland;

<sup>3</sup>Department of Pediatrics; Yong Loo Lin School of Medicine; National University of Singapore; Singapore; <sup>4</sup>KK Women's and Children's Hospital; Singapore;

<sup>5</sup>Department of Obstetrics and Gynecology; Yong Loo Lin School of Medicine; National University of Singapore; Singapore; <sup>6</sup>MRC Lifecourse Epidemiology Unit and NIHR Southampton Biomedical Research Centre; University of Southampton and University Hospital Southampton NHS Foundation Trust; Southampton, United Kingdom;

<sup>†</sup>Current address: Vishuo BioMedical Pte Ltd; Teletech Park; Singapore

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\*Correspondence to: Joanna D Holbrook; Email: Joanna\_Holbrook@sics.a-star.edu.sg

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**T**he gut of the human neonate is colonized rapidly after birth from an early sparse and highly distinct microbiota to a more adult-like and convergent state, within 1 to 3 years. The progression of colonizing bacterial species is non-random. During the first months of life several shifts commonly occur in the species prevalent in our guts. Although the sequential progression of these species is remarkably consistent across individuals and geographies, there is inter-individual variation in the rate of progression. Our study and others suggest that the rate is influenced by environmental factors, and influences our future health. In this article, we review our recent contribution to cataloging the developing infant gut microbiota alongside other important recent studies. We suggest testable hypotheses that arise from this synthesis.

The fetus is almost sterile during the gestational period,<sup>1</sup> and birth starts a process of colonization by microorganisms. How the gut is colonized is of interest for at least three reasons. Firstly, the make-up of the gut microbiota has been causally linked with metabolic health and disease.<sup>2–4</sup> Secondly, microbiota may be a mechanism for developmental programming;<sup>5</sup> while the infant gut microbial composition does not necessarily predict the make-up of the adult gut microbiota,<sup>6</sup>

it does affect the infant's developing immune system and metabolism.<sup>7,8</sup> Finally, the gut microbiota can be sampled non-invasively, albeit with some loss of accuracy and complexity, by studying fecal matter.<sup>9</sup>

Yatsunenکو et al,<sup>10</sup> among others found that the fecal microbiome varies strongly but predictably with age over the first few years of life. They also found that this progression with age was essentially consistent across Venezuela, Malawi and USA, although there were some strong differences in microbiota content across geographies. In a fascinating study of premature infants in a neonatal intensive care ward designed to minimize sources of bacterial exposure, La Rosa et al,<sup>11</sup> found that the progression of microbial species found in fecal samples followed a similar pattern to that described before and included discrete stages where particular phyla dominated. They found that extrinsic factors “influenced the pace, but not the sequence, of progression.” These studies might suggest that intrinsic factors such as gut maturity are driving the progression. External environmental factors such as delivery mode, antibiotic exposure and feeding style strongly affect the infant fecal microbiota,<sup>12</sup> but maybe only the rate of its establishment.

In our recent publication,<sup>13</sup> we found that 75 newborn babies from the Singaporean GUSTO birth cohort went through a progression of gut microbiota acquisition, similar to the studies

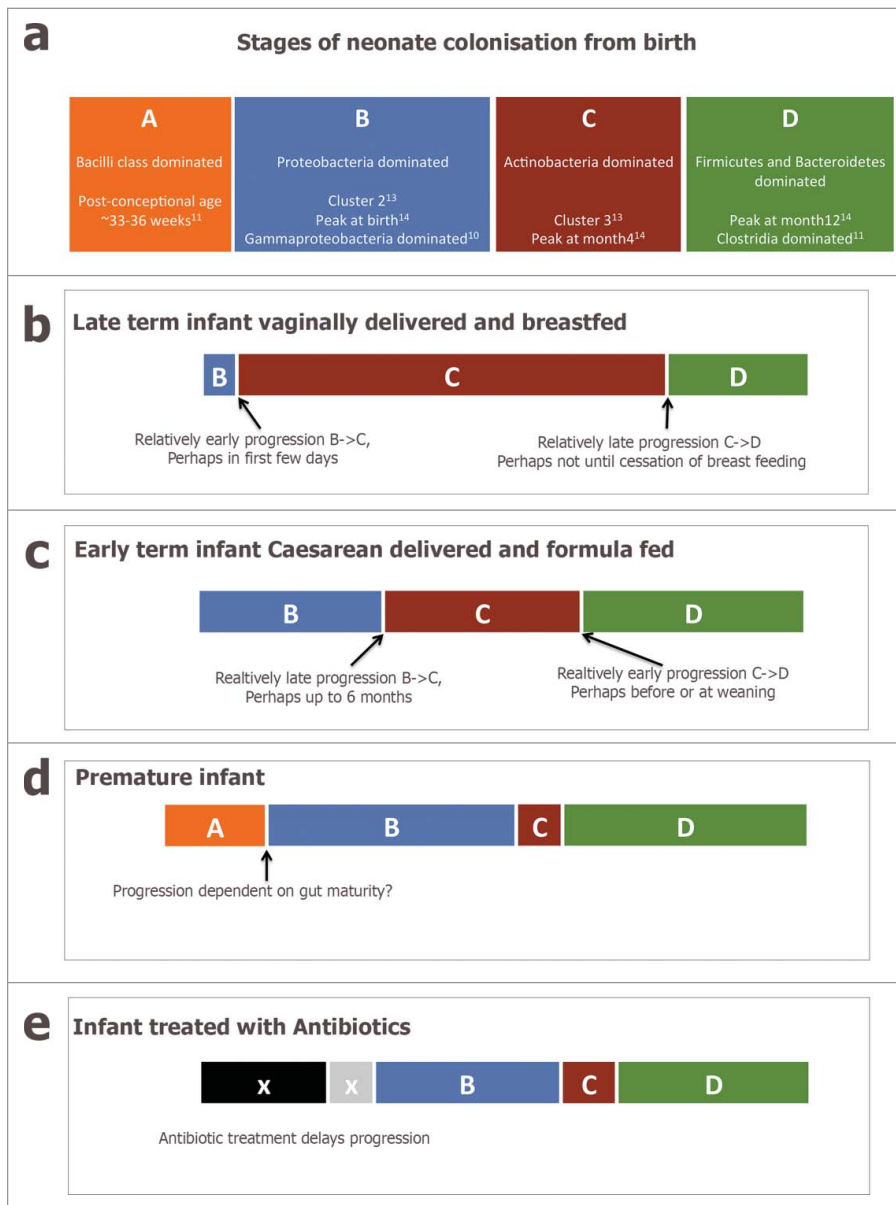
previously mentioned and others. We used 16S rRNA pyrosequencing of stool samples collected at ages 3 days, 3 weeks, 3 months and 6 months and classified the fecal microbiota by non-supervised clustering; over this period the majority of infants progressed from a “cluster-2” fecal microbial profile (high in Enterobacteriaceae and Klebsiella) to a “cluster-3” fecal microbial profile (high in Bifidobacteria and Collinsella). In our study, 88% of the babies reached a cluster-3 classified profile by age 6 months, but there was substantial

inter-individual variation in the rate at which it was reached. At day 3 26% of the neonates already had fecal microbiota which classified as cluster-3, while 1% of the babies only reached cluster-3 by month 6; the remaining reached there at week 3 (35%) or month 3 (26%). Most studies of infants that studied inter-individual variation have reported differences between individuals of the same age. However, we postulated that the rate of microbiota acquisition is an important source of inter-individual variation.

Subsequent to the publication of our article, Bäckhed et al,<sup>14</sup> published a comprehensive study of the fecal microbiota of 98 Swedish infants sampled at birth, month 4 and month 12 along with fecal samples from their mothers. Crucially, they employed metagenomic sequencing, a much more quantitative and comprehensive methodology, which allows identification of bacterial species and genes, and so can describe functional capacity. Both Bäckhed et al,<sup>14</sup> and our own study,<sup>13</sup> found that fecal samples were more similar within time-points than within individuals.

Bäckhed et al,<sup>14</sup> also described a shift in the fecal microbiota over time. Their Figure 1 showed microbiota within a week of birth very similar to our cluster 2 microbiota, high in Enterobacteriaceae species such as Escherichia/Shigella and in Streptococcus. Species characteristic of cluster 3 in our study,<sup>13</sup> such as Bifidobacterium and Collinsella, peaked in relative abundance at month 4 in Bäckhed’s data. The Swedish data showed a further large transition between the month 4 and month 12 samples. Our data did not extend past month 6 and so did not include this shift to the Bacteroidetes and Firmicutes species documented by Bäckhed et al.

Figure 1a presents a simplified schematic of what is known about the progression of species in the infant fecal microbiota over time. The Bacilli class dominated stage A is found chiefly in premature infants, as documented by La Rosa et al.,<sup>11</sup> and rarely in other infants. Stage B dominated by Proteobacteria and termed Cluster 2 in our study, seems to exist from birth or soon after in term infants; Bäckhed et al.,<sup>14</sup> also observed this profile in samples at birth, La Rosa et al.,<sup>11</sup> observed it at post-conceptual ages 32–34 weeks, and many other studies have also reported a similar profile.<sup>12</sup> Progression from Stage B to Stage C (Actinobacteria dominated, called Cluster 3 in our study) occurred between birth and month 6 in our study.<sup>13</sup> The species characteristic of Cluster 3, e.g. Bifidobacterium, peaked in relative abundance in the samples at month 4 in Bäckhed et al.<sup>14</sup> It was also the most frequent profile observed in fecal samples of 24 Canadian infants at 4 months of age.<sup>15</sup> Interestingly it was absent from the study



**Figure 1.** (a) schematic diagram representing a simplified view of the progression of the infant fecal microbiota across the first year of life, incorporating Stages (A–D). (b–e) present putative modifications to the progression caused by intrinsic and extrinsic factors.

of premature infants by La Rosa et al.<sup>11</sup> Another dramatic shift from Actinobacteria to Firmicutes and Bacteroidetes occurs between month 4 and years 1-3 in term infants,<sup>14,16</sup> and perhaps earlier in premature infants.<sup>11</sup> Stage D represents a more adult-like microbiota.<sup>10,12,14</sup>

Our thesis is that population shifts in the core microbiota are developmental-stage specific. Our study and others suggest that extrinsic factors primarily contribute to inter-individual diversity in rate of progression. Furthermore, the rate of progression influences health later in life.

In both our study,<sup>13</sup> and Bäckhed et al.<sup>14</sup> there was a profound effect of delivery mode on the infant gut microbiota, which was most marked at early time-points and decreased at later ages. In Bäckhed et al (2015) the infants born by Caesarean section had much sparser early fecal microbiota, which tended to contain species usually found on adult skin, while the fecal microbiota of the vaginally delivered infants closely resembled the fecal microbiota of their mothers. Even at the 12-month time-point the Bacteroidetes phylum was less abundant in infants born by Caesarean section. We also found that Bacteroidetes was absent from infants born at Caesarean section at all time-points, although we note that it was variably present in the vaginally born infants. These findings echo those of Jakobsson et al.<sup>17</sup> Bäckhed et al.,<sup>14</sup> found strong evidence for vertical transmission of *Bacteroides fragilis* and *Bacteroides thetaiotaomicron* among other species in vaginally delivered infants.

So, delivery mode affects the infant microbiota in cross-sectional comparisons, i.e. when comparing infants at the same age. However, this apparent difference in microbiota composition could be explained by differences in rate of progression through the same stages. In our study those infants who were vaginally delivered tended to reach the Actinobacteria dominated Cluster-3/ Stage C earlier. It is difficult to determine if the rate of progression from Stage B to Stage C is affected by delivery mode in the data of Bäckhed et al.,<sup>14</sup> but it is possible to observe in their Figure 1 that many of the vaginally delivered babies already had high levels of Bifidobacterium and Collinsella as neonates.

Bäckhed et al.<sup>14</sup> used the random forest approach, which was developed by Subramanian et al.<sup>18</sup> to describe delay in the normal development of the gut microbiota in malnourished children. They found that babies born by Caesarean section (or fed formula) had fecal microbiota that were characterized as older than their chronological age at 4 months. At first appearance this is in contrast to our conclusion,<sup>13</sup> which proposed babies born by Caesarean tended to reach Cluster 3 (Stage C) later than vaginally delivered infants. However, Cluster 3 represents a month 6 pre-weaning microbiota and not the Stage D weaned microbiota that represents maturity (and on which the random forests algorithm was trained) in the Bäckhed et al. data. Perhaps the Caesarean delivered infants spend a relatively compressed time at Stage C but instead convert relatively rapidly from Stage B to Stage D (Fig. 1c).

Premature infants have repeatedly been shown to have a markedly different gut microbiota to term infants,<sup>19</sup> with a bacilli dominated phase,<sup>11</sup> delayed or missed acquisition of Bifidobacteria,<sup>11,20,21</sup> and earlier acquisition of Firmicutes,<sup>11</sup> (Fig. 1d). Our study was unusual in showing a difference in the acquisition of gut microbiota across the normal range of gestational ages. All the infants in our study were born at term but those born at 38 weeks tended to reach Cluster 3 later than those born at 39 or more weeks' gestation. Indeed, the median gestation for those who reached Cluster 3 at day 3 was 39.2 weeks as compared to 38.5 weeks for those who reached Cluster 3 at month 6. Overall, a difference in gestational duration of about a week was associated with a lag in gut microbiota acquisition measured in months.

Others have suggested that host physiology drives the differences in gut microbiota between premature and term infants.<sup>16,22</sup> In particular a defective mucin barrier especially MUC2 may lead to unbalanced microbiota.<sup>23,24</sup> Perhaps subtle immaturity of the gut in the early term infants is enough to delay microbiota acquisition. However cause and effect are difficult to unpick: Athalye-Jape et al. showed that administering probiotics can speed up gut maturation in premature

infants,<sup>25</sup> suggesting that the gut microbiome can influence physiology and vice-versa. Administering IGF1 in utero may speed development of the guts of premature infants,<sup>26</sup> and it would be telling if this also affected the gut microbiota.

The most dramatic shift in the Bäckhed et al.<sup>14</sup> data is between the months 4 and 12, Stage C to Stage D. The month 12 microbiota of the infants is more similar to the adult profiles of their mothers than to either the birth or month 4 profiles and led the authors to speculate that infants could reach a “steady-state” profile at about one year old, similar but a bit earlier to the 1-3 y time-point proposed by Yatsunenko et al.<sup>10</sup> The introduction of solid food (weaning) usually within 4 and 12 months of life has been previously implicated in this shift,<sup>16</sup> but Bäckhed et al.<sup>14</sup> emphasize the role of breast-feeding. Infants that were still breastfed at age 12 months tended to have fecal microbiota containing the Actinobacterial species seen at 4 months of age and characteristic of Stage C. Also, as mentioned above the machine learning approach employed by Bäckhed et al.<sup>14</sup> determined that formula fed infants tended to have a profile that was characterized as “older” than that of breast fed infants at the birth and 4-month time-points. Therefore, breastfeeding seems to delay the progression to Stage D, even after introduction of solid food.

In summary, the timing of the transition between Stages B and C is affected by delivery mode and gestational duration. Similarly, the timing of the transition between Stage C and D seems to also be influenced by mode of delivery and more importantly the intake of breast milk and solid food. An obvious following question is: does the timing matter?

Cox et al.<sup>27</sup> showed in mice that disruption of the early microbiota (by a low dose of penicillin limited to gestation and infancy) could enhance the effect of diet in inducing obesity in adulthood. In humans too, early treatment with antibiotics is associated with later adiposity.<sup>28,29</sup> Taking antibiotics delays the progression of the gut microbiota,<sup>30,31</sup> (Fig. 1e). Antibiotic usage at the end of gestation and during early infancy correlates with increased body

size in childhood,<sup>32</sup> and risk of being overweight.<sup>33</sup>

Babies born prematurely show a lag in the proper colonization of the gut,<sup>31</sup> and less time at Bifidobacteria dominated Stage C;<sup>20</sup> they are also at higher risk for obesity later in life.<sup>34,35</sup> Similarly individuals born by Caesarean section have a slower rate of progression to Stage C,<sup>13,14</sup> and are also at higher risk for obesity.<sup>36,37</sup> However, longer duration of breastfeeding is associated with longer at Stage C,<sup>14</sup> and is protective of obesity.<sup>36,38</sup> In our study,<sup>13</sup> those infants which reached Cluster 3 (aka Stage C) later were more likely to be of median adiposity at 18 months old, while those who got there later were more likely to be significantly underweight at age 18 months. Those who had high levels of the Firmicutes genera *Streptococcus* at the month 6 time-point (characteristic of Stage D and so presumably these infants paused at Stage C for a shorter time), had a higher than average adiposity gain between birth and 18 months of age. Previous cross-sectional studies have found lower Bifidobacterium levels in fecal samples from children younger than 6 months correlates with future obesity at 7 and 10 y of age, compared to normal weight controls.<sup>39,40</sup>

Many researchers are asking if changing the early microbiota by pre- or probiotics, could affect later phenotype and perhaps prevent obesity.<sup>41</sup> In agreement with the hypothesis presented here, others have suggested that infants born at earlier gestations,<sup>42</sup> or by Caesarean section,<sup>43</sup> would benefit from intervention to get them to Stage C faster and keep them there longer.<sup>44,45</sup>

Our present understanding of the progression of the infant gut microbiota is crude. **Figure 1** represents a testable hypothesis but is oversimplified and most likely incorrect in detail. More studies sampling infants at more frequent time-points are needed to build a better and more complete picture. Employing the type of metagenomic approach exemplified by Bäckhed et al,<sup>14</sup> is not only more quantitative but starts to provide insights into the functional capacity of the microbiota. Even more informative would be meta-transcriptomic approaches that provide

information of which genes are expressed in the microbiome.

The following is a short-listing of some of the most pressing questions:

- (1) There are almost certainly more stages we have yet been able to capture - what are they?
- (2) Are the stages discrete or are they more gradual than our sparse data suggests at present?
- (3) Do all infants progress through all stages or do some infants miss some stages completely?
- (4) What are the intrinsic and extrinsic factors that influence speed of progression at different points? How do they interact?
- (5) Several studies have documented the influence of the maternal microbiome and vertical transmission,<sup>46</sup> is this how factors such as delivery mode affect rate of progression?
- (6) Is it beneficial to progress through some stages fast and others more slowly? For instance, is it beneficial for future metabolic health to progress through Stage C slowly?
- (7) It is interesting to note that the recovery of the childhood and adult gut microbiota after diarrhea has also been observed to be orderly and reproducible.<sup>47,48</sup> If as suggested by the authors, intrinsic factors govern the progression: are there similarities within individuals between the original process of colonization and the re-colonization that occurs after diarrhea?

#### Disclosure of Potential Conflicts of Interest

OS, CNB, WB, BB and HB are employees of the Nestlé Research Center, a commercial entity that aims to enhance the quality of consumers' lives through nutrition, health and wellness. Nestlé is active in research into prebiotics and probiotics and produces infant formula. KMG and YSC have received reimbursement for speaking at conferences sponsored by companies selling nutritional products, and are part of an academic consortium that has received research funding from Abbott Nutrition, Nestec and Danone. The other authors have no

potential conflicts of interest. A patent in a related field has been submitted (No. 14181275.0).

#### References

1. Aagaard K, Ma J, Antony KM, Ganu R, Petrosino J, Versalovic J. The placenta harbors a unique microbiome. *Sci Transl Med* 2014; 6:237ra65; PMID:24848255
2. Gao X, Jia R, Xie L, Kuang L, Feng L, Wan C. Obesity in school-aged children and its correlation with Gut E. coli and Bifidobacteria: a case-control study. *BMC Pediatr* 2015; 15:64; PMID:26024884; <http://dx.doi.org/10.1186/s12887-015-0384-x>
3. Karl JP, Fu X, Wang X, Zhao Y, Shen J, Zhang C, Wolfe BE, Saltzman E, Zhao L, Booth SL. Fecal menaquinone profiles of overweight adults are associated with gut microbiota composition during a gut microbiota-targeted dietary intervention. *Am J Clin Nutr* 2015; 102(1):84-93; PMID:26016865
4. Zhong CY, Sun WW, Ma Y, Zhu H, Yang P, Wei H, Zeng BH, Zhang Q, Liu Y, Li WX, et al. Microbiota prevents cholesterol loss from the body by regulating host gene expression in mice. *Sci Rep* 2015; 5:10512; PMID:26015368; <http://dx.doi.org/10.1038/srep10512>
5. Putignani L, Del Chierico F, Petrucca A, Vernocchi P, Dallapiccola B. The human gut microbiota: a dynamic interplay with the host from birth to senescence settled during childhood. *Pediatr Res* 2014; 76:2-10; PMID:24732106; <http://dx.doi.org/10.1038/pr.2014.49>
6. Knights D, Ward TL, McKinlay CE, Miller H, Gonzalez A, McDonald D, Knight R. Rethinking "enterotypes." *Cell Host Microbe* 2014; 16:433-7; PMID:25299329; <http://dx.doi.org/10.1016/j.chom.2014.09.013>
7. Collado MC, Cernada M, Bauerl C, Vento M, Perez-Martinez G. Microbial ecology and host-microbiota interactions during early life stages. *Gut Microbes* 2012; 3:352-65; PMID:22743759; <http://dx.doi.org/10.4161/gmic.21215>
8. Kerr CA, Grice DM, Tran CD, Bauer DC, Li D, Hendry P, Hannan GN. Early life events influence whole-of-life metabolic health via gut microflora and gut permeability. *Crit Rev Microbiol* 2014; 41(3):326-40; PMID:24645635
9. Romano-Keeler J, Moore DJ, Wang C, Brucker RM, Fonnesebeck C, Slaughter JC, Li H, Curran DP, Meng S, Correa H, et al. Early life establishment of site-specific microbial communities in the gut. *Gut Microbes* 2014; 5:192-201; PMID:24637795; <http://dx.doi.org/10.4161/gmic.28442>
10. Yatsunenkov T, Rey FE, Manary MJ, Trehan I, Dominguez-Bello MG, Contreras M, Magris M, Hidalgo G, Baldassano RN, Anokhin AP, et al. Human gut microbiome viewed across age and geography. *Nature* 2012; 486:222-7; PMID:22699611
11. La Rosa PS, Warner BB, Zhou Y, Weinstock GM, Sodergren E, Hall-Moore CM, Stevens HJ, Bennett WE, Jr., Shaikh N, Linneman LA, et al. Patterned progression of bacterial populations in the premature infant gut. *Proc Natl Acad Sci USA* 2014; 111:12522-7; PMID:25114261; <http://dx.doi.org/10.1073/pnas.1409497111>
12. Matamoros S, Gras-Leguen C, Le Vacon F, Potel G, de La Cochetiere MF. Development of intestinal microbiota in infants and its impact on health. *Trends Microbiol* 2013; 21:167-73; PMID:23332725; <http://dx.doi.org/10.1016/j.tim.2012.12.001>
13. Dogra S, Sakwinska O, Soh SE, Ngom-Bru C, Bruck WM, Berger B, Brussow H, Lee YS, Yap F, Chong YS, et al. Dynamics of infant gut microbiota are influenced by delivery mode and gestational duration and are associated with subsequent adiposity. *MBio* 2015; 6:e02419-14; PMID:25650398

14. Backhed F, Roswall J, Peng Y, Feng Q, Jia H, Kovatcheva-Datchary P, Li Y, Xia Y, Xie H, Zhong H, et al. Dynamics and Stabilization of the Human Gut Microbiome during the First Year of Life. *Cell Host Microbe* 2015; 17:690-703; PMID:25974306; <http://dx.doi.org/10.1016/j.chom.2015.04.004>
15. Azad MB, Konya T, Maughan H, Guttman DS, Field CJ, Chari RS, Sears MR, Becker AB, Scott JA, Kozyrskiy AL, et al. Gut microbiota of healthy Canadian infants: profiles by mode of delivery and infant diet at 4 months. *CMAJ* 2013; 185:385-94; PMID:23401405
16. Gritz EC, Bhandari V. The human neonatal gut microbiome: a brief review. *Front Pediatr* 2015; 3:17; PMID:25798435
17. Jakobsson HE, Abrahamsson TR, Jenmalm MC, Harris K, Quince C, Jernberg C, Bjorksten B, Engstrand L, Andersson AF. Decreased gut microbiota diversity, delayed Bacteroidetes colonisation and reduced Th1 responses in infants delivered by caesarean section. *Gut* 2014; 63:559-66; PMID:23926244; <http://dx.doi.org/10.1136/gutjnl-2012-303249>
18. Subramanian S, Huq S, Yatsunenkov T, Haque R, Mahfuz M, Alam MA, Benezra A, DeStefano J, Meier MF, Muegge BD, et al. Persistent gut microbiota immaturity in malnourished Bangladeshi children. *Nature* 2014; 510:417-21; PMID:24896187
19. Groer MW, Luciano AA, Dishaw LJ, Ashmeade TL, Miller E, Gilbert JA. Development of the preterm infant gut microbiome: a research priority. *Microbiome* 2014; 2:38; PMID:25332768; <http://dx.doi.org/10.1186/2049-2618-2-38>
20. Butel MJ, Suau A, Campeotto F, Magne F, Aires J, Ferraris L, Kalach N, Leroux B, Dupont C. Conditions of bifidobacterial colonization in preterm infants: a prospective analysis. *J Pediatr Gastroenterol Nutr* 2007; 44:577-82; PMID:17460489; <http://dx.doi.org/10.1097/MPG.0b013e3180406b20>
21. Collado MC, Cernada M, Neu J, Perez-Martinez G, Gormaz M, Vento M. Factors influencing gastrointestinal tract and microbiota immune interaction in preterm infants. *Pediatr Res* 2015; 77:726-31; PMID:25760550; <http://dx.doi.org/10.1038/pr.2015.54>
22. Stockinger S, Hornef MW, Chassin C. Establishment of intestinal homeostasis during the neonatal period. *Cell Mol Life Sci* 2011; 68:3699-712; PMID:21952827; <http://dx.doi.org/10.1007/s00018-011-0831-2>
23. Elson CO, Alexander KL. Host-microbiota interactions in the intestine. *Dig Dis* 2015; 33:131-6; PMID:25925913; <http://dx.doi.org/10.1159/000369534>
24. Cobo ER, Kissoon-Singh V, Moreau F, Chadee K. Colonic MUC2 mucin regulates the expression and antimicrobial activity of beta-defensin 2. *Mucosal Immunol* 2015; PMID:25921338; <http://dx.doi.org/10.1038/mi.2015.27>
25. Athalye-Jape G, Deshpande G, Rao S, Patole S. Benefits of probiotics on enteral nutrition in preterm neonates: a systematic review. *Am J Clin Nutr* 2014; 100:1508-19; <http://dx.doi.org/10.3945/ajcn.114.092551>
26. Kimble RM, Breier BH, Gluckman PD, Harding JE. Enteral IGF-I enhances fetal growth and gastrointestinal development in oesophageal ligated fetal sheep. *J Endocrinol* 1999; 162:227-35; PMID:10425460; <http://dx.doi.org/10.1677/joe.0.1620227>
27. Cox LM, Yamanishi S, Sohn J, Alekseyenko AV, Leung JM, Cho I, Kim SG, Li H, Gao Z, Mahana D, et al. Altering the intestinal microbiota during a critical developmental window has lasting metabolic consequences. *Cell* 2014; 158:705-21; PMID:25126780; <http://dx.doi.org/10.1016/j.cell.2014.05.052>
28. Trasande L, Blustein J, Liu M, Corwin E, Cox LM, Blaser MJ. Infant antibiotic exposures and early-life body mass. *Int J Obes* 2013; 37:16-23; PMID:22907693; <http://dx.doi.org/10.1038/ijo.2012.132>
29. Azad MB, Bridgman SL, Becker AB, Kozyrskiy AL. Infant antibiotic exposure and the development of childhood overweight and central adiposity. *Int J Obes* 2014; 38:1290-8; <http://dx.doi.org/10.1038/ijo.2014.119>
30. Dardas M, Gill SR, Grier A, Pryhuber GS, Gill AL, Lee YH, Guillet R. The impact of postnatal antibiotics on the preterm intestinal microbiome. *Pediatr Res* 2014; 76:150-8; PMID:24819377; <http://dx.doi.org/10.1038/pr.2014.69>
31. Arboleya S, Sanchez B, Milani C, Duranti S, Solis G, Fernandez N, de los Reyes-Gavilan CG, Ventura M, Margolles A, Guimonde M. Intestinal microbiota development in preterm neonates and effect of perinatal antibiotics. *J Pediatr* 2015; 166:538-44; <http://dx.doi.org/10.1016/j.jpeds.2014.09.041>
32. Gough EK, Moodie EE, Prendergast AJ, Johnson SM, Humphrey JH, Stoltzfus RJ, Walker AS, Trehan I, Gibb DM, Goto R, et al. The impact of antibiotics on growth in children in low and middle income countries: systematic review and meta-analysis of randomised controlled trials. *Bmj* 2014; 348:g2267; PMID:24735883; <http://dx.doi.org/10.1136/bmj.g2267>
33. Ajslev TA, Andersen CS, Gamborg M, Sorensen TI, Jess T. Childhood overweight after establishment of the gut microbiota: the role of delivery mode, pre-pregnancy weight and early administration of antibiotics. *Int J Obesity* 2011; 35:522-9; <http://dx.doi.org/10.1038/ijo.2011.27>
34. Lane RH. Fetal programming, epigenetics, and adult onset disease. *Clin Perinatol* 2014; 41:815-31; PMID:25459776; <http://dx.doi.org/10.1016/j.clp.2014.08.006>
35. Bayman E, Drake AJ, Piyasena C. Prematurity and programming of cardiovascular disease risk: a future challenge for public health? *Arch Dis Child Fetal Neonatal Ed* 2014; 99:F510-4; PMID:25135955; <http://dx.doi.org/10.1136/archdischild-2014-306742>
36. Portela DS, Vieira TO, Matos SM, de Oliveira NF, Vieira GO. Maternal obesity, environmental factors, cesarean delivery and breastfeeding as determinants of overweight and obesity in children: results from a cohort. *BMC Pregnancy Childbirth* 2015; 15:94; PMID:25884808; <http://dx.doi.org/10.1186/s12884-015-0518-z>
37. Kuhle S, Tong OS, Woolcott CG. Association between caesarean section and childhood obesity: a systematic review and meta-analysis. *Obes Rev* 2015; 16:295-303; PMID:25752886; <http://dx.doi.org/10.1111/obr.12267>
38. Zhu Y, Hernandez LM, Dong Y, Himes JH, Hirschfeld S, Forman MR. Longer breastfeeding duration reduces the positive relationships among gestational weight gain, birth weight and childhood anthropometrics. *J Epidemiol Community Health* 2015; 69(7):632-8; PMID:25680365
39. Kalliomaki M, Collado MC, Salminen S, Isolauri E. Early differences in fecal microbiota composition in children may predict overweight. *Am J Clinical Nutr* 2008; 87:534-8
40. Luoto R, Kalliomaki M, Laitinen K, Delzenne NM, Cani PD, Salminen S, Isolauri E. Initial dietary and microbiological environments deviate in normal-weight compared to overweight children at 10 years of age. *J Pediatr Gastroenterol Nutr* 2011; 52:90-5; <http://dx.doi.org/10.1097/MPG.0b013e3181f3457f>
41. Subramanian S, Blanton LV, Frese SA, Charbonneau M, Mills DA, Gordon JI. Cultivating Healthy Growth and Nutrition through the Gut Microbiota. *Cell* 2015; 161:36-48; PMID:25815983; <http://dx.doi.org/10.1016/j.cell.2015.03.013>
42. Dutta S, Ray P, Narang A. Comparison of Stool Colonization in Premature Infants by Three Dose Regimes of a Probiotic Combination: A Randomized Controlled Trial. *Am J Perinatol* 2014; 32(8):733-40; PMID:25519197
43. Miniello VL, Colasanto A, Cristofori F, Diaferio L, Ficele L, Lieggi MS, Santoiemma V, Francavilla R. Gut microbiota biomodulators, when the stork comes by the scalpel. *Clin Chim Acta* 2015; PMID:25668229; <http://dx.doi.org/10.1016/j.cca.2015.01.022>
44. Pacheco AR, Barile D, Underwood MA, Mills DA. The impact of the milk glycomiome on the neonate gut microbiota. *Annu Rev Anim Biosci* 2015; 3:419-45; PMID:25387230; <http://dx.doi.org/10.1146/annurev-animal-022114-111112>
45. Arboleya S, Bahrami B, Macfarlane S, Guimonde M, Macfarlane GT, de Los Reyes-Gavilan CG. Production of immune response mediators by HT-29 intestinal cell-lines in the presence of Bifidobacterium-treated infant microbiota. *Benef Microbes* 2015; 6(4):1-10; PMID:25691102
46. Mueller NT, Bakacs E, Combellick J, Grigoryan Z, Dominguez-Bello MG. The infant microbiome development: mom matters. *Trends Mol Med* 2015; 21:109-17; PMID:25578246; <http://dx.doi.org/10.1016/j.molmed.2014.12.002>
47. David LA, Weil A, Ryan ET, Calderwood SB, Harris JB, Chowdhury F, Begum Y, Qadri F, LaRocque RC, Turnbaugh PJ. Gut microbial succession follows acute secretory diarrhea in humans. *mBio* 2015; 6:e00381-15
48. Hsiao A, Ahmed AM, Subramanian S, Griffin NW, Drewry LL, Petri WA, Jr., Haque R, Ahmed T, Gordon JI. Members of the human gut microbiota involved in recovery from *Vibrio cholerae* infection. *Nat* 2014; 515:423-6; PMID:25231861; <http://dx.doi.org/10.1038/nature13738>