

1 **Synergizing food waste management and microalgae biorefinery for bioenergy production:**
2 **recent advance on direct and indirect conversion pathway**

3 Adityas Agung Ramandani ¹, Sze Ying Lee ^{2,*}, Anet Režek Jambrak ³, Wei-Hsin Chen ^{4,5,6}, Jun
4 Wei Lim ⁷, Kuan Shiong Khoo ^{1,*}

5

6 ¹ Algae Bioseparation Research Laboratory, Department of Chemical Engineering and Materials
7 Science, Yuan Ze University, Taoyuan, Taiwan

8 ² Singapore Institute of Food and Biotechnology Innovation (SIFBI), Agency for Science,
9 Technology and Research (A*STAR), 31 Biopolis Way, Nanos, Singapore 138669, Singapore

10 ³ Faculty of Food Technology and Biotechnology, University of Zagreb, Zagreb, Croatia

11 ⁴ Department of Aeronautics and Astronautics, National Cheng Kung University, Tainan 701,
12 Taiwan

13 ⁵ Research Center for Smart Sustainable Circular Economy, Tunghai University, Taichung 407,
14 Taiwan

15 ⁶ Department of Mechanical Engineering, National Chin-Yi University of Technology, Taichung
16 411, Taiwan

17 ⁷ HICoE-Centre for Biofuel and Biochemical Research, Institute of Self-Sustainable Building,
18 Department of Fundamental and Applied Sciences, Universiti Teknologi PETRONAS, 32610
19 Seri Iskandar, Perak Darul Ridzuan, Malaysia

20

21 ***Corresponding authors:**

22 **Dr. Sze Ying Lee**

23 Singapore Institute of Food and Biotechnology Innovation (SIFBI), Agency for Science,
24 Technology and Research (A*STAR), 31 Biopolis Way, Nanos, Singapore 138669, Singapore

25 Email: lee_sze_ying@sifbi.a-star.edu.sg

26

27 **Assistant Professor ChM. Dr. Kuan Shiong Khoo, Ph.D**

28 Algae Bioseparation Research Laboratory, Department of Chemical Engineering and Materials
29 Science, Yuan Ze University, Taoyuan, Taiwan

30 Email: kuanshiong.khoo@saturn.yzu.edu.tw or kuanshiong.khoo@hotmail.com

31 Abbreviations

| | |
|------|-------------------------------|
| ABE | Acetone-butanol-ethanol |
| C/N | Carbon to nitrogen |
| CNT | Carbon nanotubes |
| COD | Chemical oxygen demand |
| DCW | Dry cell weight |
| FAME | Fatty acid methyl ester |
| GHG | Greenhouse gasses |
| HHV | Higher heating value |
| LHV | Lower heating value |
| MNP | Magnetic nanoparticle |
| PAC | Powdered activated carbon |
| SDGs | Sustainable development goals |
| VS | Volatile solid |

32

33 **Abstract**

34 Food waste is a persistent global environmental issue that contributes to global warming and
35 climate change by releasing significant amounts of greenhouse gases as it decomposes in landfills.
36 Converting food waste into bioenergy could serve as a sustainable solution. Direct conversion of
37 food waste through methods like anaerobic digestion, fermentation, and pyrolysis, produces
38 diverse energy products. Besides, the indirect approach involving the cultivation of microalgae on
39 food waste offers a sustainable approach, owing to the ability of microalgae to assimilate nutrient-
40 rich components such as nitrogen, phosphorus, and carbon from food waste, converting them into
41 valuable biomass. The microalgae biomass can be further transformed into biofuels, offering a
42 dual benefit of waste management and energy production. This review work provides a holistic
43 review of food waste-to-energy conversion methods, both direct and indirect, evaluating their
44 feasibility and benefits. Specifically, a comprehensive analysis of using microalgae grown on food
45 waste for biofuel production is provided, offering insights into how the method could effectively
46 address both environmental and energy crises, and contribute to sustainable development goals.
47 Future research should focus on enhancing conversion process efficiency, optimizing microalgae
48 bioprocess, and scaling-up these technologies to industrial levels, while addressing challenges such
49 as economic feasibility, supportive policies, and robust supply chains. Collaboration among
50 industry, academia, and government is essential to boost the global implementation of these
51 technologies.

52

53 **Keywords:** Bioenergy; food waste; sustainable development goals; biotransformation; third-
54 generation biofuels; microalgae

55 **1. Introduction**

56 Food waste, one of the largest global waste streams at 1.05 billion ton in 2022 from retail, food
57 service, and household sectors, poses significant environmental and economic challenges due to
58 its unsustainable disposal practices [1]. The rapid growth of the world's population drives up the
59 food demand, and at the same time, substantial amounts of food waste are being generated from
60 farms, grocery stores, food service sectors, and households. It is estimated that one-third of all
61 food produced globally is lost or wasted in the food supply chain [2]. By mass, food waste consists
62 of 62% fruit and vegetables, 11% meat, 11% dairy, and 16% bakery products, with 39% classified
63 as edible and 61% inedible [1]. Food waste contributes significantly to the municipal waste, with
64 over 60% of the trash found in a restaurant's bin being food waste [3]. Most of the generated food
65 waste ends up in landfill; however, in terms of net greenhouse gas (GHG) emission, it appears to
66 be the worst disposal option [4]. The GHG released from landfilled food waste is remarkably high
67 compared to those generated from other wastes such as paper, wood, and yard trimmings, releasing
68 around 2708 kg carbon dioxide equivalent (CO₂e) ton⁻¹ of dry food waste [5]. There are several
69 alternatives to landfill disposal. Composting has been promoted with its advantages of converting
70 food waste into fertilizer with the reduced GHG emissions as low as about 926 kg CO₂e ton⁻¹ of
71 dry food waste, which was 38 – 84% lower than landfilling [6]. However, composting requires
72 maintenance, including regular watering and aeration, and large-scale implementation faces
73 challenges such as limited land availability in urban areas and the need for odor control measures
74 to mitigate unpleasant smells.

75 In this context, the transformation of food waste into bioenergy presents a promising
76 alternative for effective waste management and energy recovery. Food waste sorting is currently
77 practiced in many countries, facilitating its valorization into bioenergy. However, the least

78 developed countries still lack sorting systems for waste streams and do not implement effective
79 conversion pathways for its valuation. Several conversion methods are available. Anaerobic
80 digestion, a method with its initial use on decomposing sewage sludge at wastewater treatment
81 facilities, is increasingly popular and frequently used to dispose food waste and generate renewable
82 energy (biogas, mostly methane) and nutrient rich-digestate. It was reported that the anaerobic
83 digestion is favored among various food waste disposal options in view of its relatively lower
84 GHG emissions compared to composting and landfills [4]. However, food waste from different
85 sources, along with uncontrolled mixing with agricultural feedstocks, influence methane yield,
86 since the biodegradability varies significantly. Higher levels of proteins and lipids in food waste
87 lead to increased concentrations of ammonia and sulfide, which inhibit the metabolism of
88 anaerobic consortia [7]. Apart from biomethane, another renewable energy source (biohydrogen),
89 can be produced from anaerobic fermentation of food waste. In fact, biohydrogen is an
90 intermediate product of anaerobic digestion, and lately there has been a gaining interest in
91 optimizing its production [8]. Furthermore, pyrolysis represents an efficient thermochemical
92 conversion option for converting food waste into bio-oil, biochar, and bio-syngas [9]. It can be
93 used to treat food waste with large amounts of solid impurities and even the recalcitrant organic
94 residues generated from anaerobic digestion of food waste [9].

95 Alternatively, processed solid food waste and/or liquid food waste such as waste cooling
96 oil can be served as a low-cost nutrient source to grow microorganisms such as bacteria, fungi,
97 yeast, and microalgae. Microalgae are an attractive choice due to their rapid growth rates, high
98 biomass productivity, and ability to utilize a wide range of food waste substrates for nutrient
99 uptake. Their ability to metabolize diverse waste-derived nutrients, including glucose, nitrogen,
100 and phosphorous, enables the conversion of food waste into valuable biomass composition (i.e.

101 carbohydrates, proteins, and lipids) [10]. These food waste-derived substrates provide an important
102 resource for reducing the costs associated with synthetic cultivation medium. Furthermore,
103 microalgae also contribute significantly to the circular bioeconomy by transforming food waste
104 nutrients into biofuels, thereby supporting sustainable development goals. Microalgae-based
105 biofuels have been progressed over the past decade to replace petroleum-based energy source. The
106 waste nutrients from food waste can be upcycled as an alternative culture medium for the
107 production of microalgal biomass, and its whole biomass and isolated products can be further
108 exploited to produce various types of biofuels (e.g. biodiesel, bioethanol, and biogas). For instance,
109 food waste hydrolysate rich in glucose and fructose was served as the carbon source to support the
110 heterotrophic growth of *Auxenochlorella protothecoides*, achieving a biomass yield of 0.346 g g⁻¹
111 sugar and a lipid yield of 0.216 g g⁻¹ sugar [11]. By applying the proposed integrated method, 1
112 kg of food waste produced 248 g of biodiesel [11]. Microalgae do not require arable land or fresh
113 water and can be fully utilized for biofuel production, making them an ideal solution for food waste
114 valorization. Furthermore, microalgae contribute to carbon sequestration, helping mitigate the
115 effects of climate change. As research continues to optimize upstream processes and enhance
116 downstream processing, microalgae could play a pivotal role in advancing sustainable, low-carbon
117 energy solutions. Promising results have been frequently reported in the relevant studies, implying
118 that microalgae cultivation using food waste nutrients holds potential to revolutionize biofuel
119 industries [12]. Regardless of the different conversion methods used, process economics and
120 profitability remain critical factors in determining the sustainability of these technologies. Cost
121 competitiveness is crucial for biofuels, and their market viability is highly influenced by the price
122 of fossil fuels. A recent study demonstrated that the total operational cost for cultivating *Chlorella*
123 *vulgaris* FSP-E can be reduced to $\$2.15 \times 10^5$ by using industrial biscuit waste powder as the

124 cultivation medium, compared to $\$3.78 \times 10^6$ when using the BG-11 medium [13]. Despite that,
125 the successful implementation of food waste valorization to energy requires careful examination
126 of advancements in conversion technologies

127 The novelty of this review lies in its holistic approach, evaluating the advancements of both
128 direct conversion (e.g., anaerobic digestion, fermentation, and pyrolysis) and indirect
129 biotransformation (e.g., microbial biomass cultivation using food waste) methods for turning food
130 waste into bioenergy. The potential for integrating various direct conversion methods and the
131 interaction between direct and indirect approaches is examined. The integration could maximize
132 resource utilization, reduce waste, and enhance the efficiency of biofuel production while
133 contributing to environmental sustainability and circular bioeconomy. Additionally, pretreatment
134 of food waste prior to conversion processes is essential to ensure process efficiency, since it
135 enhances the availability of organic materials. This work aims to provide insights into the effective
136 food waste management protocols that can be implemented on a large-scale to valorize food waste
137 sustainably. Finally, the challenges and perspectives are outlined.

138 **2. Food waste and their pretreatment for valorization**

139 Different forms of food waste influence the choice of conversion technologies due to their varying
140 composition, moisture content, and biodegradability. Solid food waste can be processed and
141 converted into biogas rich in methane through anaerobic digestion, biohydrogen via fermentation,
142 and bio-oil, biochar, and pyrolysis gas through pyrolysis (**Figure 1**). Besides, liquid food waste
143 (e.g., waste cooking oil) and lipid holds within solid food waste can be converted into biodiesel
144 through esterification and transesterification [14]. In another indirect approach, processed solid and
145 liquid food waste can be used as feedstocks for microbial fermentation, supporting the growth of

146 microbial cells that are then converted into various energy products, including biodiesel and
147 bioethanol/biobutanol (**Figure 2**).

148 Solid food waste needs to undergo multi-step pretreatments such as mechanical, physical,
149 chemical, and biological methods. Mechanical pretreatment (e.g., blending, shredding, grinding,
150 and pulverizing) is indispensable to reduce the size of food waste solids, making them more
151 accessible to the subsequent treatment. Size reduction and sieving pretreatment are needed to
152 ensure the efficiency of anaerobic digestion and prevent clogging of the digestion system. The
153 selection of pretreatment is dependent on the nature of food waste and the requirements of the
154 waste-to-energy conversion techniques. For wet anaerobic digestion, water was added into blended
155 food waste and the feedstock was processed in the form of dilute organic slurry with typically a
156 low total solid of < 15% [15]. In contrast, the feedstock for dry anaerobic digestion with > 15% of
157 total solids was prepared by drying blended food waste at 105 °C in hot air oven for 24 h [16]. In
158 dry fermentation, the colonization of biomass by anaerobic consortia is slow due to the absence of
159 additional liquid in the chambers, which limits their metabolism and subsequently reduces biogas
160 production. The disintegration of biowaste (e.g., shockwave pretreatment) enhances the
161 fermentation process, and can be implemented either before the biowaste enters dry anaerobic
162 fermentation or during later stages when the anaerobic consortia have exhausted the easily
163 biodegradable sources of organic matter [17]. Similar to anaerobic digestion, pretreatment is
164 indispensable to increase pyrolysis product yields, e.g., solid food waste requires extensive drying
165 to remove its water content prior to pyrolysis [18]. It was also reported that the food waste
166 pretreated by torrefaction before pyrolysis yielded high-quality bio-oils with the increased heating
167 values ranging from 31.51 – 34.34 MJ kg⁻¹ compared to that of obtained from unpretreated
168 feedstocks (27.69 – 31.58 MJ kg⁻¹) [19]. High-quality bio-oil is typically characterized by its low

169 moisture content, low oxygen content, and high energy density, making it more suitable for use as
170 a fuel.

171 Additionally, complex compounds within food waste are broken down into simple molecules
172 using physical, chemical, and biological pretreatment, allowing an effective nutrient assimilation
173 by microorganisms. Physical method like thermal treatment is effective for certain types of food
174 waste but often come with high cost and moderate environmental impacts due to its high energy
175 consumption. On the other hand, chemical pretreatment, though highly efficient in breaking down
176 complex compounds, is also costly and can lead to potential environmental issues due to chemical
177 residues. Another milder pretreatment, which involves microorganisms or enzymes, is more
178 environmentally friendly but may require longer processing time and specific conditions for
179 optimal efficiency. Generally, pretreatment steps increase overall production cost and energy
180 consumption considering the buildup and operational costs of infrastructure and facility, and the
181 resources required. Therefore, it is important to carefully evaluate the benefits and drawbacks of
182 pretreatment processing before implementing it on an industrial scale. Alternatively, by-products
183 like liquid digestate discharged from anaerobic digestion, which is rich in nitrogen and phosphorus,
184 can be also utilized for microorganism cultivation to optimize cost-effectiveness and sustainability
185 in the waste-to-energy conversion platform [20]. Apart from that, hydrothermal technologies
186 present promising alternatives for valorizing food waste with high moisture content, since it
187 eliminates the need for pre-drying wet materials, which in turn reducing energy costs and lowering
188 operational expenses.

189 **3. Direct conversion of food waste to biofuels**

190 Conversion of wasted food to biofuels is one of the best options to divert from landfills. Landfilling
191 of food waste is illegal in most developed countries, and this ban advocates for the use of more

192 effective methods to reduce residual waste and advance toward a circular economy. Anaerobic
193 digesters and pyrolysis facilities can be located in highly populated urban areas, whereby the
194 energy content of food waste is recovered, and the volume is significantly reduced, without the
195 need for a long-distance transportation. Recent studies are summarized in **Table 1**, showcasing
196 various conversion technologies used to obtain different biofuel products. Each method has its
197 own pros and cons. For example, fermentation offers high energy efficiency and low
198 environmental impact but requires additional processing steps for biofuel production. Meanwhile,
199 pyrolysis has advantages in energy recovery and waste volume reduction but it is energy-intensive
200 [21].

201 **3.1 Production of biomethane**

202 Anaerobic digestion involves the use of bacteria to break down organic matters in the absence of
203 oxygen and produces biogas rich in biomethane, which can be served as an energy source. The
204 digestate, the residual material left after the digestion process, can be used as fertilizer or further
205 processed using other techniques such as pyrolysis. Anaerobic digestion encompasses several main
206 phases, including hydrolysis, acidification, acetogenesis, and methanogenesis, which play critical
207 roles in affecting the efficiency of biogas production. Previously conducted research have
208 validated the need of pretreatment step to improve the hydrolysis of organic matters in food waste.
209 Food waste is usually complex and constituted of high amounts of fats (i.e., 3 – 6% on wet basis)
210 that impede the effective hydrolysis of other substrates, and thus pretreatment is applied to improve
211 hydrolytic rate and efficiency. Chen et al. [22] showed that the food waste pretreated with alkali
212 enhanced the solubilization of its organic matters and achieved a high yield of biogas at 829 mL
213 g⁻¹ volatile solid (VS) with 65.5% of methane. Another recent study also showed that food waste,
214 which has been pretreated using the subcritical water hydrolysis method, was mixed with pulp

215 wastewater at a volatile solids (VS) ratio of 3:1 for anaerobic digestion, resulting in an increased
216 biogas yield of 807 mL g⁻¹ VS [23].

217 It is a common practice to mix food waste with other co-digested materials, such as manure
218 and agricultural residues, to enhance the overall efficiency of anaerobic digestion while balancing
219 the economic performance of the process. Anaerobic digestion of food waste often incorporates
220 cow dung, since cow dung contains many types of bacteria that can act as decomposers in the
221 process. It was reported that the mixed liquid tofu waste and cow dung at a percentage composition
222 of 75:25, in the presence of 5% of EM-4 biocatalyst, produced biogas containing > 90% of methane
223 and with a higher heating value (HHV) and a lower heating value (LHV) of 50,149 and 46,909 kJ
224 kg⁻¹, respectively [24]. Besides, anaerobic co-digestion of canteen food waste and sewage
225 treatment plant sludge, at its optimum ratio of 3:1, yielded high biogas production at 280 – 860 mL
226 g⁻¹ VS [25]. Another work demonstrated the use of catering waste mixed with the pretreated
227 parthenium weed for biogas production [26]. These reports implied that food waste is a potential
228 feedstock for enhanced biogas production. Recently, the use of additives such as biochar [27] and
229 conductive materials like magnetite nanoparticle (MNP), carbon nanotubes (CNT), and powdered
230 activated carbon (PAC) [28] have shown to improve the process efficiency of anaerobic digestion.
231 The addition of biochar was proven to promote the acetogenesis and hydrogenotrophic
232 methanogenesis processes in a continuous two-stage anaerobic digestion system, and improved
233 the energy recovery efficiency by 25 – 71% [27]. On the other hand, conductive materials
234 displayed their abilities to improve production of biohydrogen and biomethane in a two-stage
235 anaerobic fermentation. Particularly, PAC facilitated high removal efficiency of chemical oxygen
236 demand (COD) and increased yield of biomethane in the second stage of anaerobic fermentation
237 [28].

238 **3.2 Production of biohydrogen**

239 Food waste has a high content of carbohydrates, making it a suitable feedstock for hydrogen
240 production. Typically, the hydrogen-producing potential of carbohydrate is relatively greater than
241 lipid and protein, and thus hydrogen yield is frequently reported on a hexose basis. The theoretical
242 maximum yield of hydrogen generated from hexose is 4 mol of H₂ mol⁻¹ hexose, but the actual
243 yield is usually < 50% of theoretical value due to thermodynamic constraints [29]. The hydrogen
244 yields reported in literature are generally below 3 mol of H₂ mol⁻¹ hexose [30]. Various techniques
245 are available for producing biohydrogen from food waste, which can be broadly categorized into
246 light fermentation and dark fermentation. Light fermentation utilizes photosynthetic bacteria that
247 converts sunlight into energy to produce biohydrogen, while dark fermentation involves the
248 adoption of anaerobic bacteria like *Clostridium* species for producing biohydrogen in the absence
249 of light [31]. The latter technique is considered more superior owing to its rapid hydrogen
250 production and low operation cost without the need for a light source. The selection of pretreatment
251 technique is dependent on the composition of food waste. Common pretreatment techniques used
252 include physical (e.g., thermal and sonication), chemical (e.g., acids and alkalis), and biological
253 (e.g., use of enzymes and microorganisms) [32]. Kim et al. [33] found that the fermentative
254 hydrogen production from food waste without the addition of inoculum was possible given that
255 the food waste has undergone appropriate pretreatment. Amongst the pretreatment methods
256 including acid, alkali, and thermal treatments, food waste that was thermally treated at 90 °C for
257 20 min obtained the highest yield (i.e., 1.65 mol H₂ mol⁻¹ hexose) [33]. On the other hand, Lee et
258 al. [34] demonstrated an alternative method of using a glucose-adapted species, *Thermococcus*
259 *onnurineus* NA1, which has undergone adaptive evolution process, to produce hydrogen using
260 potato peel waste through simultaneous saccharification and fermentation.

261 Apart from that, there are several key parameters influencing the efficiency of hydrogen
262 generation through fermentation, including feedstock composition, operating pH, temperature, and
263 agitation speed [35]. The variations in composition and quality of food waste increase the
264 complexity level of pretreatment, demanding a careful process optimization to ensure consistent
265 and efficient product conversion. Xue et al. [36] found that a highly concentrated food waste
266 contributed to the strong substrate acidification at the beginning of fermentation, inactivating non-
267 hydrogen producing bacteria and consequently enhanced the yield up to 120.78 mL H₂ g⁻¹ VS.
268 Another report highlighted that the pH and total solid content in food waste were also two
269 important factors in lactate-driven dark fermentation that allowed a concomitant production of
270 hydrogen and medium-chain carboxylic acids such as caproate [37]. The study showed that a more
271 neutral pH (at 6.5) and an increment of solid concentration (at 8% total solids) favored to hydrogen
272 production using dark fermentation, yielding about 103.4 NmL H₂ g⁻¹ VS and a simultaneous
273 production of 8.7 g L⁻¹ of caproate [37]. Overall, hydrogen production from waste seems viable,
274 with the recent advancements in waste refining making it more cost-effective at €5.4 (100 km)⁻¹
275 compared to fossil fuels at €15.6 (100 km)⁻¹ [38]. Nonetheless, its infrastructure for everyday
276 mobility is still in the early stage globally.

277 **3.3 Production of bio-oil, biochar and bio-syngas**

278 Conversion of food waste into liquid (bio-oil), non-condensable gaseous (bio-syngas), and solid
279 residue (biochar) products involves a thermal decomposition process that occurs in the absence of
280 oxygen. In the process, solid food waste is heated at the elevated temperature to break down into
281 its constituent parts. The resulting biochar can be used as a soil amendment, while bio-oil can be
282 further processed into liquid fuels, and bio-syngas can be used for power generation. Solid food
283 waste is pretreated before pyrolysis process to remove its water content. Pyrolysis of food waste

284 with the assistance of microwave irradiation have been examined. Kadlimatti et al. [39]
285 demonstrated that the production of bio-oil from microwave-assisted pyrolysis of food waste with
286 a yield of 30.24% under the optimized conditions, and the bio-oil has a heating value of 23.94 MJ
287 kg^{-1} . Nonetheless, the bio-oil obtained is unsuitable for use as biofuel owing to its high contents
288 of water and nitrogenated compounds [39]. Likewise, the bio-oil produced from the pyrolysis of
289 waste cooking oil reported in another work also required further processing because of its high
290 acidity index of 126.8 mg KOH g^{-1} oil and high viscosity of 8.95 cSt, though it has a high HHV at
291 8843 kg Kcal $^{-1}$ [40]. Besides, the work showed the bio-oil yield can be optimized to reach 80wt%
292 with the use of final pyrolysis temperature of 800 °C with a heating rate of 15 °C min^{-1} [40].

293 In addition to food waste, pyrolysis can be used to treat solid digestate produced from anaerobic
294 digestion of food waste. A recent study by Zhao and co-workers [41] compared the quality of the
295 products yielded from the pyrolysis of food waste and food waste solid digestate, respectively, and
296 found that the pyrolytic gas derived from food waste has an increased LHV by 8% compared to
297 that of acquired from food waste solid digestate. Nonetheless, the biochar obtained from food
298 waste solid digestate possessed a specific surface area of about 10-fold greater than that of attained
299 from food waste, suggesting its potential use as an adsorption material. Moreover, the pyrolytic oil
300 from food waste solid digestate displayed the characteristics that are suitable for use as fuel. This
301 work highlighted the benefits of combining pyrolysis and anaerobic digestion [41]. Furthermore,
302 pyrolysis can be coupled with gasification for enhanced hydrogen fraction in the syngas produced.
303 It was reported that the hydrogen content increased to 66.7% in syngas using *in-situ* gasification
304 after pyrolysis [42]. In contrast to pyrolysis, advanced methods such as HTL process wet biomass
305 at elevated pressures and temperatures to produce energy-dense biocrude oil and a nutrient-rich
306 aqueous phase product. It was reported that HTL of simulated food waste yielded approximately

307 30wt% of bio-oil with HHV of 35 MJ kg⁻¹ [43]. The optimal conditions for achieving high yield
308 of biocrude oil was at temperatures close to the critical point (i.e., 374 °C) and pressures sufficient
309 to keep water in a liquid or dense (> 0.4 g cm⁻³) supercritical phase [43].

310 **4. Indirect transformation of food waste to biofuels through microorganism cultivation**

311 Indirect transformation involves the use of microorganisms that are cultivated on nutrient-rich food
312 waste hydrolysate to produce bioenergy (**Figure 2**). Various microorganism species, including
313 bacteria, fungi, yeast, and microalgae, have displayed wide adaptability to grow on food waste
314 hydrolysate. They can accumulate lipids or carbohydrates within the cells which can be processed
315 into biofuels. Alternatively, the whole and spent microbial biomass can be served as feedstock in
316 conversion processes for turning it into energy products. Integrating microbial technology with
317 food waste management for biofuel production presents a sustainable alternative to traditional
318 energy systems (e.g., fossil fuels and direct food waste conversion to bioenergy). This approach
319 enhances the economic feasibility of microbial-based biofuels, since relying on synthetic growth
320 media is unsustainable for large-scale microalgae biorefineries due to the high costs associated
321 with the chemicals.

322 **4.1 Bacteria, fungi, and yeast grown on food waste**

323 Oil-producing microorganisms or known as oleaginous microorganisms for those accumulate >
324 20% of lipids on a dry cell weight (DCW) basis, are the alternative lipid sources for biodiesel
325 production. Bacteria (e.g., *Bacillus subtilis*, *Rhodococcus opacus*, and *Gordonia* sp.), fungi (e.g.,
326 *Aspergillus oryzae*, *Mortierella isabellina*, and *Humicola lanuginosa*) and yeast (e.g., *Yarrowia*
327 *lipolytica*, *Cryptococcus curvatus*, and *Lipomyces starkeyi*) species have been recognized for their
328 high lipid contents. For instance, *Rhodospiridium toruloides* grown on catering food waste
329 hydrolysate achieved a high biomass productivity of 32.9 g L⁻¹, with 36.4% lipid content [44]. The

330 biodiesel produced from the lipids extracted met the satisfactory performance according to the
331 European standard EN 14214 [44]. Similarly, food waste saccharified liquid can be used to support
332 cultivation of *R. toruloides* without any additional nutrient supplementation, resulting in a biomass
333 production of 12.09 g L⁻¹ with 52.7% lipids [45]. The promising results evidently suggested that
334 biodiesel production through culturing oleaginous microorganisms on food waste is feasible and
335 holds industrial application prospects.

336 **4.2 Microalgae grown on food waste**

337 In addition, microalgae, key feedstocks for third- and fourth-generation biofuels, display wide
338 adaptability to thrive on food waste hydrolysate. The macro- and micro-nutrients within food waste
339 can be upcycled through nutrient assimilation by microalgae cells and transformed into useful
340 compounds such as lipid, carbohydrate, and protein, which can be converted to value-added energy
341 products (**Figure 3**). Microalgae are classified into prokaryotic (e.g., *Cyanophyta* and
342 *Prochlorophyta*) and eukaryotic (e.g., *Glaucophyta*, *Rhodophyta*, *Heterokontophyta*, *Haptophyta*,
343 *Cryptophyta*, *Dinophyta*, *Euglenophyta*, *Chlorarachniophyta*, and *Chlorophyta*) microorganisms.
344 The high lipid content of microalgae, ranging from 20 – 70%, makes them potential feedstocks for
345 the production of biodiesel [46]. For examples, *Chlorella sorokiniana* HS1, *Tetraselmis* sp., and
346 *Chlorella protothecoides* have been reported to yield lipid contents of 37.5, 51.7, and 35.3%,
347 respectively [47-49]. Triglycerides are used in the biodiesel production because of their higher
348 fatty acid content (>50%) and the absence of impurities (e.g., phosphate, sulfur, and nitrogen) that
349 interfere with the final product [50]. It was reported that *Schizochytrium mangrovei* can grow well
350 on food waste hydrolysate in heterotrophic cultivation, achieving a growth rate that was twice than
351 that of growth on conventional medium [51]. Under controlled nutrient-deficient conditions, *S.*
352 *mangrovei* grown on food waste hydrolysate accumulated 32% lipids, 39% carbohydrates, and

353 13% proteins, with the majority of fatty acids present in the lipids are suitable for biodiesel
354 production. In the study, the canteen food waste was hydrolyzed using fungal *Aspergillus*
355 *awamori* and *Aspergillus oryzae* for 24 h to produce hydrolysate rich in glucose (31.9 g (100 g)⁻¹
356 dry food waste), free amino nitrogen (0.28 g (100 g)⁻¹ dry food waste), and phosphate (0.38 g (100
357 g)⁻¹ dry food waste) [51]. Besides, enzymatically hydrolyzed food waste was reported to support
358 growth of *C. sorokiniana*, with the biomass produced containing 26.4% lipids and 37.8% proteins
359 [52]. Another recent report also demonstrated that the food waste hydrolysate with an initial
360 glucose concentration of 10 g L⁻¹ was successfully used as culture medium in the mixotrophic
361 cultivation of *Chlorella* sp. GY-H4 [53]. The resultant microalgal lipid had a high triglyceride
362 content (28.8%), and the biodiesel produced exhibited improved quality, e.g., with an iodine value
363 of 70 g I₂ (100 g)⁻¹, which was much lower than the maximum limit of 120 g I₂ (100 g)⁻¹ [53].

364 Alongside with lipid synthesis, microalgae accumulate carbohydrate inside their chloroplasts
365 (for eukaryotes) and cytosols (for prokaryotes), which are mainly composed of monosaccharide
366 and polysaccharide for structural and metabolic purposes [54]. Carbohydrate can be converted to
367 bioenergy (e.g., bioethanol, biobutanol, and biogas) through microbial fermentation [55]. Like
368 lipids, carbohydrate content in microalgal cell and its sugar profile can be enhanced by shifting its
369 cell metabolism towards carbohydrate accumulation through upstream cultivation strategies. For
370 example, carbohydrate content of *C. vulgaris* increased by 72% when cultured in modified BG-11
371 medium compared to standard medium, reaching 58% [56]. Ajala and Alexander [57] reported that
372 high carbohydrate contents of 29, 38, and 33% were found in *C. vulgaris*, *Scenedesmus obliquus*,
373 and *Oocystis minuta*, respectively. *Chlamydomonas* sp. also achieved 50.5 and 19.0% of
374 carbohydrate and lipid, which can be used to produce bioethanol and biodiesel, respectively [58].
375 *S. mangrovei* and *C. pyrenoidosa* cultivated on food waste hydrolysate accumulated up to 38.9 and

376 31.0% carbohydrates, respectively, under nutrient-restricted culture conditions [51]. Additionally,
377 it was suggested that the depleted food waste hydrolysate used for microalgae cultivation, which
378 was still rich in sugars and organic acids (e.g., lactic acid) and not metabolized by *C. sorokiniana*,
379 can be used as the feedstock for anaerobic digestion for producing biogas [52].

380 **5. Biorefinery of microalgae for biofuel production**

381 Given the significant potential of microalgae for biofuel production, advancing their conversion
382 technologies is essential to improve process efficiency, enabling better utilization of microalgae as
383 a sustainable energy source. **Figure 4** explicitly details different biorefinery routes for microalgal
384 biomass to produce various biofuel products, and the related studies are summarized in **Table 2**.

385 **5.1 Production of biodiesel**

386 Microalgal lipid can be converted to biodiesel via ex-situ transesterification, which is a two-step
387 approach involving cell pretreatment and lipid extraction prior to the transesterification process.
388 Conventional lipid extraction strategies such as Bligh and Dyer, Folch, and Soxhlet methods are
389 available and the advanced techniques using ionic liquid, deep eutectic solvent, and switchable
390 solvent have also been explored [59]. Besides, direct transesterification (in-situ) allows lipid
391 extraction and transesterification to occur in a single step within the same reaction vessel. Karimi
392 et al. [60] reported that the *in-situ* transesterification achieved 99% conversion of bio-oil from
393 *Nannochloropsis* to biodiesel, and the biodiesel yield was about 8% of DCW. The remarkable
394 conversion efficiency obtained using in-situ transesterification method indicates its potential for
395 streamlining biodiesel production process by reducing the number of processing steps involved.
396 Likewise, Yasin et al. [61] demonstrated that direct transesterification achieved a higher fatty acid
397 methyl ester (FAME) yield than two-step method. However dry biomass is preferred because of
398 the presence of water in wet biomass led to losses of FAME composition in direct

399 transesterification process. Therefore, more research works are needed to improve the application
400 of direct transesterification method on wet biomass, to reduce the need for the energy-intensive
401 drying.

402 **5.2 Production of bioethanol and biobutanol**

403 Carbohydrate-rich biomass appears as an alternative renewable feedstock for acetone–butanol–
404 ethanol (ABE) fermentation to produce alcohol-based biofuels such as butanol and ethanol. The
405 processes involve acidogenesis and solventogenesis [61]. During acidogenesis, microalgal
406 biomass is fermented to produce organic acids, such as acetic acid and butyric acid, and the process
407 is accompanied with a drop of pH levels [62]. These organic acids are then converted into
408 biobutanol through solventogenesis, in which the solvent (e.g., acetone, butane and ethanol) and
409 bacteria (e.g., *Clostridium acetobutylicum*) are added to ferment the acids produced [62]. Al-
410 Shorgani et al. [63] reported that the ABE fermentation using *C. acetobutylicum* YM1 increased
411 the yield of biobutanol from 13.82 to 21.71 g L⁻¹. Apart from that, Condor et al. [64] showed that
412 the bioethanol of 0.18, 0.17 and 0.2 g g⁻¹ biomass were obtained from *C. vulgaris* FSP-E with the
413 use of *Saccharomyces cerevisiae* FAY-1, *S. cerevisiae* FAY-2, and *Zymomonas mobilis*,
414 respectively. The findings highlight the importance of selecting suitable strains for maximizing
415 bioethanol production, indicating that the choice of fermentation microorganism is critical for
416 optimizing yield. Besides, advanced pretreatment methods can significantly enhance the
417 accessibility of carbohydrates in microalgae biomass, achieving improved fermentation efficiency
418 and overall yield. For instance, Yirgu et al. [65] reported that the bioethanol yield of 0.08 g g⁻¹
419 biomass was achieved in a 24-h fermentation process after the microwave-assisted acid hydrolysis
420 pretreatment.

421 **5.3 Production of biogas**

422 Anaerobic digestion can be used to produce biogas from dry and wet microalgal biomass, and even
423 microalgal biomass residues after lipid and pigment extraction [66]. Microalgal biomass is
424 converted into gaseous methane and carbon dioxide in the methanogenic process after the
425 acetogenic stage. The multi-step approach maximizes the efficiency of biogas production from
426 microalgal biomass. Vargas-Estrada et al. [66] identified the factors influencing the yield,
427 including pH and pretreatment used (e.g., thermal, thermochemical, and enzymatic methods). The
428 findings of Xiao et al. [67] ascertained the need of pretreatment to break down organic compounds
429 and increase the availability of substrate for methanogenic bacteria. The biogas produced from
430 microalgal biomass pretreated with solar-driven hydrothermal process displayed a relatively high
431 exergy efficiency (40.85%) compared to that of obtained from the biomass without any
432 pretreatment (26.2%) [67]. It was also found that the organic loading rate and temperature during
433 anaerobic digestion affected the yield of biogas [68]. These findings highlight the need for a robust
434 process design to maximize energy recovery from the anaerobic digestion of microalgal biomass.

435 **5.4 Production of biohydrogen**

436 The approaches for the production of biohydrogen from microalgae include bio-photolysis and
437 fermentation [69]. Direct bio-photolysis involves the conversion of water molecules into hydrogen
438 and oxygen with the use of sunlight and photosynthetic microorganisms, whereas indirect bio-
439 photolysis utilizes synthetic catalysts or enzymes to increase process efficiency. By combining the
440 natural abilities of microorganism and catalyst, indirect bio-photolysis is capable to optimize
441 hydrogen production and overcomes the limitations of natural system. Bio-photolysis can be
442 facilitated by harnessing the capabilities of microalgae and cyanobacteria since these organisms
443 are capable of carrying out the reaction efficiently. A recent study showed that a multispecies algae-

444 bacteria consortium formed by *Chlamydomonas reinhardtii* and *Microbacterium forte* sp. yielded
445 sustainable production of up to 313 mL H₂ L⁻¹ for 17 days [70].

446 On the other hand, fermentation can be grouped into photo- and dark-fermentation. Photo-
447 fermentation uses light energy to convert organic matters into hydrogen using photosynthetic
448 bacteria (e.g., purple non-sulfur bacteria, purple sulfur bacteria, and green sulfur bacteria). In
449 contrast, dark fermentation relies on the use of anaerobic bacteria, particularly thermophile and
450 mesophile microorganisms, to break down organic matters and produces hydrogen alongside with
451 byproducts such as lactic, acetic, propionic, butyric, and ethanol. Sharma et al. [71] reviewed the
452 approaches of hydrogen generation from organic waste and concluded that there are still
453 technological challenges such as the pretreatment method, operational cost, and gaseous hydrogen
454 storage. Besides, high carbohydrate-accumulating microalgal strains are sought after considering
455 the inevitable loss of carbohydrate during pretreatment [72].

456 **5.5 Production of bio-oil, biochar, and bio-syngas**

457 Unlike some other biomass types, microalgal biomass does not require extensive pretreatment
458 before pyrolysis. This is due to the unique composition of microalgae, which typically contains
459 higher amounts of lipid, protein, and carbohydrate, but lower amounts of lignin compared to
460 lignocellulosic biomass. Bio-oil, biochar, and bio-syngas can be produced from microalgal
461 biomass via conventional and advanced pyrolysis. Conventional pyrolysis can be grouped into
462 slow, intermediate, and fast, which are mainly characterized by the operating temperature range,
463 which are 400 – 500, 400 – 650, and 850 – 1250 °C, respectively [73]. The classification of
464 pyrolysis methods underscores the significance of temperature in determining the yield and quality
465 of bio-products. For instance, the yield of bio-oil obtained from *C. vulgaris* increased from 30.9 to
466 48.1% when the temperature was raised from 300 to 700°C [74].

467 Recently, advanced pyrolysis technologies such as catalytic pyrolysis (i.e., with the use of
468 catalyst), co-pyrolysis (i.e., mixture of two or more biomass), hydro-pyrolysis (i.e., with the use
469 of pressurized hydrogen and hydrogen-based technologies) and microwave-assisted pyrolysis have
470 been investigated [75]. Ferreira and Soares Dias [76] reported that the different catalysts were
471 added in the pyrolysis of *C. vulgaris*, such as Na_2CO_3 and MgCO_3 , yielding bio-oils of 30.6 and
472 42.8%, respectively. The addition of catalyst in the pyrolysis process significantly affects the
473 process efficiency. Further research is needed to optimize the selection of appropriate catalyst for
474 maximum bio-oil yield.

475 **6. Research needs and way forward**

476 With the growing awareness of sustainability, there has been a significant increase in the number
477 of developed countries recognizing the importance of waste-to-energy conversion technologies.
478 Cost is one of the most critical considerations for implementing food waste-to-energy systems.
479 The large capital investment in infrastructure and technology, along with the ongoing costs for
480 operation and maintenance, must be balanced against the environmental and economic benefits of
481 food waste-to-energy conversion system to ensure long-term feasibility and promote large-scale
482 adoption. The realization of food waste-to-energy plants on both national and international scales
483 needs a careful techno-economic assessment, along with the environmental evaluation. Huiru et
484 al. [77] analyzed the economic viability of an anaerobic digestion plant fed with food waste
485 collected from 29 canteens in a China university, and a simple payback period of 7.8 years was
486 obtained. The results showed that the project's economic feasibility was highly dependent on the
487 electricity price and initial plant investment, with a 10% change of these two parameters potentially
488 extending the equity payback to a maximum of 12.4 years [77]. Another recent techno-economic
489 analysis by Chen et al. [78] supported the use of anaerobic digestion to treat city-scale food waste

490 for biogas production, with most of the studied pathways showed positive net present values in the
491 baseline scenario. In addition, the proposed approaches were found to be advantageous in reducing
492 carbon footprints over the landfills. Other direct conversion method for biohydrogen and bio-oil
493 production also showed potential for economic gain [79, 80] and should be further explored. It is
494 noteworthy to highlight that the concurrent production of additional valuable products alongside
495 biofuels from food waste can significantly improve economic viability of the process by
496 diversifying revenue streams and reducing reliance on a single product. This has been validated in
497 the techno-economic analysis of Rajendran and Han [81] that the plant has a short project payback
498 period of 7.31 years for the integrated production of polyhydroxyalkanoates and biofuels such as
499 biohydrogen, bioethanol, and 2,3-butanediol from food waste.

500 On the other hand, microalgae-based bioenergy from food waste has been initiated since
501 2000 and continue to develop in recent years (**Figure 5**). The approach seems promising but
502 significant challenges to commercialization remain. Although microalgae cultivation on food
503 waste greatly reduces the chemical expenses associated with synthetic media, the overall
504 production costs remain high due to the expenses of waste pretreatment processes, the setup and
505 operation of photobioreactors, and the biorefinery process. Open pond systems offer the benefits
506 of reduced construction and maintenance costs particularly for large-scale cultivation, but they
507 face challenges such as contamination and difficulty to maintain optimal productivity due to
508 uncontrollable environmental conditions. It was reported that the cultivation method affects the
509 production cost substantially, with the FAME from *Spirulina* produced from an open pond system
510 costs around \$4 kg⁻¹, while those cultivated using a solar-lit photobioreactor costs approximately
511 \$25 kg⁻¹ [82]. Similar to direct conversion methods, the generation of co-products is particularly
512 beneficial in microalgae-based bioenergy production, creating extra revenue that can be used to

513 improve the final selling price of the fuel, as highlighted by Dutta et al. [83] in their techno-
514 economic analysis. The calculated minimum fuel selling price was improved from \$10.55 to \$4.35
515 gasoline gallon equivalent (GGE)⁻¹ by incorporating additional processes, including fermentation
516 and anaerobic digestion, to generate co-products such as bioethanol, biogas, and digestate along
517 with biodiesel, maximizing the valorization of microalgal biomass.

518 The integrated zero-waste microalgal biorefinery concept aims to maximize the utilization
519 of all components of microalgal biomass, thereby minimizing waste and creating a circular,
520 sustainable production process. The economic and environmental benefits of the integrated
521 microalgal biorefinery are substantial. By fully utilizing all components of the biomass, the process
522 becomes more economically viable. The extraction of microalgal lipids for biodiesel leaves behind
523 a protein-rich residue, which can be used to produce animal feed or as an ingredient in aquaculture
524 feed, reducing reliance on conventional feedstocks that contribute to deforestation and greenhouse
525 gas emissions [84]. Bioactive compounds (e.g., carotenoids, chlorophylls, and phycobiliproteins)
526 can also be extracted for use in food and pharmaceutical industries, since these products display
527 numerous health benefits. The integrated biorefinery also focuses on the production of bioethanol
528 or biobutanol from carbohydrates from microalgae through fermentation processes. Lastly, the
529 biomass residue can be used as the feedstock for anaerobic digestion to generate biogas, ensuring
530 that no biomass goes to waste. Additionally, in order to create a circular bioeconomy concept in
531 microalgae-based biofuels, by-products and waste generated should be continuously repurposed
532 to maximize resource efficiency. For instance, carbon emissions from industrial processes can be
533 fed directly into microalgal cultivation units, providing CO₂ for photosynthesis. This approach
534 contributes to reducing carbon emissions, supporting the transition to a low-carbon economy.

535 Future works should prioritize reducing microalgae production costs to address current
536 challenges and accelerate the progress of microalgae-based bioenergy. Advancements in food
537 waste pretreatment method and robust cultivation strategy to address the inconsistent nutrient
538 contents of food waste are essential to improve the efficiency and reliability of the cultivation of
539 microalgae on food waste. Additionally, developing an integrated biorefinery approach allows for
540 the simultaneous production of multiple products, which can improve overall efficiency and
541 profitability. The lipid extraction techniques for microalgae require improvement, since the use of
542 hexane is extremely problematic due to its large contribution to GHG emissions [83]. Continued
543 improvement in conversion technology is essential to enhance the effectiveness of transforming
544 microalgal biomass into bioenergy products. Integrating data-driven technologies such as machine
545 learning and artificial intelligence into food waste management for process optimization and scale-
546 up could offer promising opportunities.

547 It must be emphasized that energy is inherently a political issue, with significant social
548 consequences. Therefore, the shift to sustainable energy sources, such as microalgae-based
549 biofuels, has the potential to reshape not only environmental outcomes but also socio-political
550 dynamics by reducing dependence on fossil fuels and promoting energy security. Government
551 policies play a crucial role in supporting the scaling up of microalgae cultivation using food waste
552 for biofuel production. Subsidies for initial investments help mitigate the high costs associated
553 with infrastructure and technology, making projects more financially viable. Government
554 regulations that mandate or incentivize the use of biofuels in transportation and energy sectors
555 create market demand and ensure a stable market for biofuel producer. For example, the attempts
556 made to achieve affordable and clean energy (SGD 7) also aid in reducing GHGs and combat
557 climate change (SDG 13), while also promoting sustainable economic development (SDG 8) and

558 protecting ecosystems and biodiversity (SDGs 14 and 15). Furthermore, the establishment of
559 partnerships fosters knowledge sharing, technology transfer, and capacity building, ultimately
560 accelerating progress towards all the SDGs [85]. Baldassarre et. al [86] highlighted that the
561 research institutions need to work together with industry partners to validate the proposed solutions
562 on an industrial-scale. With these efforts, research institutions provide evidence-based solutions to
563 realize the integrated zero-waste microalgal biorefinery concept [87]. Continued development
564 efforts are crucial to overcoming existing challenges and maximizing the potential benefits of
565 sustainable biofuel production from food waste on a global scale.

566 **7. Conclusions**

567 Both direct and indirect conversion approaches contribute to food waste management and biofuel
568 production. Direct conversion methods efficiently convert food waste into biofuels but may
569 generate byproduct like sludge that requires further treatment or disposal. On the other hand, the
570 microalgae-based bioenergy platform offers dual benefits, including the recovery of food waste
571 nutrients and the generation of biomass that can be transformed into multiple bioenergy products,
572 effectively closing the loop in the food waste-to-energy cycle. However, this approach requires a
573 complex infrastructure that encompasses both upstream to downstream processes, resulting in
574 higher capital investments. An optimal technology for producing biofuels from food waste should
575 leverage both direct and indirect conversion methods by integrating them to harness their
576 complementary strengths. Continued research, technological advancements, and supportive
577 policies are crucial for realizing the full potential of these bioenergy production methods,
578 ultimately contributing to a more sustainable and resilient energy future.

579 **Acknowledgement**

580 This work was supported by the National Science and Technology Council, Taiwan (Project no.
581 NSTC112-2222-E-155-005) and the Department of Chemical Engineering and Material Science,
582 Yuan Ze University, Taiwan, under New Faculty Research Start-Up Fund Scheme (Project no:
583 303014-1 and 303014-2). The author would also like to acknowledge the support provided by
584 Research and Development (RD) Office, Yuan Ze University, Taiwan, under Assistant Teacher
585 Research Scheme (Project no: 113-HRD-07).

586

587 **References**

- 588 [1] UNEP, Think Eat Save: Tracking Progress to Halve Global Food Waste, 2024. (accessed 13
589 January 2024)
- 590 [2] FAO, Food Wastage Footprint: Impacts on Natural Resources: Summary Report, 2013.
591 <http://www.fao.org/3/i3347e/i3347e.pdf>. (accessed 29 October 2024)
- 592 [3] L. Sakaguchi, N. Pak, M.D. Potts, Tackling the issue of food waste in restaurants: Options for
593 measurement method, reduction and behavioral change, *J. Clean. Prod.* 180 (2018) 430-436.
594 <https://doi.org/10.1016/j.jclepro.2017.12.136>
- 595 [4] J.A. Moulton, S.R. Allan, C.N. Hewitt, M. Berners-Lee, Greenhouse gas emissions of food waste
596 disposal options for UK retailers, *Food Policy* 77 (2018) 50-58.
597 <https://doi.org/10.1016/j.foodpol.2018.04.003>
- 598 [5] U. Lee, J. Han, M. Wang, Evaluation of landfill gas emissions from municipal solid waste
599 landfills for the life-cycle analysis of waste-to-energy pathways, *J. Clean. Prod.* 166 (2017) 335-
600 342. <https://doi.org/10.1016/j.jclepro.2017.08.016>
- 601 [6] T. Pérez, S.E. Vergara, W.L. Silver, Assessing the climate change mitigation potential from
602 food waste composting, *Sci. Rep.* 13(1) (2023) 7608. [https://doi.org/10.1038/s41598-023-34174-](https://doi.org/10.1038/s41598-023-34174-z)
603 [z](https://doi.org/10.1038/s41598-023-34174-z)
- 604 [7] J. Maroušek, O. Strunecký, L. Kolář, M. Vochozka, M. Kopecký, A. Maroušková, J. Batt, M.
605 Poliak, M. Šoch, P. Bartoš, Advances in nutrient management make it possible to accelerate biogas
606 production and thus improve the economy of food waste processing, *Energy Sources, Part A:
607 Recovery, Util. Environ. Eff.* 46 (2020) 9379-9388.
608 <https://doi.org/10.1080/15567036.2020.1776796>

- 609 [8] H. El Bari, N. Lahboubi, S. Habchi, S. Rachidi, O. Bayssi, N. Nabil, Y. Mortezaei, R. Villa,
610 Biohydrogen production from fermentation of organic waste, storage and applications, Cleaner
611 Waste Systems 3 (2022) 100043. <https://doi.org/10.1016/j.clwas.2022.100043>
- 612 [9] A.S. Giwa, H. Xu, F. Chang, X. Zhang, N. Ali, J. Yuan, K. Wang, Pyrolysis coupled anaerobic
613 digestion process for food waste and recalcitrant residues: fundamentals, challenges, and
614 considerations, Energy Sci. Eng. 7(6) (2019) 2250-2264. <https://doi.org/10.1002/ese3.503>
- 615 [10] A.A. Ramandani, Y.-M. Sun, J.C.-W. Lan, W.-H. Chen, J.-S. Chang, N. Rachmadona, J.W.
616 Lim, K.S. Khoo, Upcycling nutrients derived from food waste via microalgae cultivation: A review
617 on impacts on cellular compounds, economy and environment analyses for achieving circular
618 bioeconomy, Biochem. Eng. J. (2024) 109454. <https://doi.org/10.1016/j.bej.2024.109454>
- 619 [11] A. Patel, K. Hružová, U. Rova, P. Christakopoulos, L. Matsakas, Sustainable biorefinery
620 concept for biofuel production through holistic valorization of food waste, Bioresour. Technol. 294
621 (2019) 122247. <https://doi.org/10.1016/j.biortech.2019.122247>
- 622 [12] Y. Kumar, S. Kaur, A. Kheto, M. Munshi, A. Sarkar, H.O. Pandey, A. Tarafdar, R. Sindhu, R.
623 Sirohi, Cultivation of microalgae on food waste: Recent advances and way forward, Bioresour.
624 Technol. 363 (2022) 127834. <https://doi.org/10.1016/j.biortech.2022.127834>
- 625 [13] A.P. Peter, X. Tan, J.Y. Lim, K.W. Chew, A.K. Koyande, P.L. Show, Environmental analysis
626 of *Chlorella vulgaris* cultivation in large scale closed system under waste nutrient source, Chem.
627 Eng. J. 433 (2022) 134254. <https://doi.org/10.1016/j.cej.2021.134254>
- 628 [14] S.K. Karmee, D. Linardi, J. Lee, C.S.K. Lin, Conversion of lipid from food waste to biodiesel,
629 Waste Manag. 41 (2015) 169-173. <https://doi.org/10.1016/j.wasman.2015.03.025>

630 [15] N. Nagao, N. Tajima, M. Kawai, C. Niwa, N. Kurosawa, T. Matsuyama, F.M. Yusoff, T. Toda,
631 Maximum organic loading rate for the single-stage wet anaerobic digestion of food waste,
632 *Bioresour. Technol.* 118 (2012) 210-218. <https://doi.org/10.1016/j.biortech.2012.05.045>

633 [16] M. Alam, M.B. Sultan, M. Mehnaz, C.S.U. Fahim, S. Hossain, A.H. Anik, Production of
634 biogas from food waste in laboratory scale dry anaerobic digester under mesophilic condition,
635 *Energy Nexus* 7 (2022) 100126. <https://doi.org/10.1016/j.nexus.2022.100126>

636 [17] V. Stehel, A. Maroušková, L. Kolář, O. Strunecký, S. Shreedhar, Advances in dry fermentation
637 extends biowaste management possibilities, *Energy Sources, Part A: Recovery, Util. Environ. Eff.*
638 42(2) (2020) 212-218. <https://doi.org/10.1080/15567036.2019.1587066>

639 [18] S.I. Okopi, J. Wang, W. Kong, Z. Yu, E.A. Ndudi, L. Che, Z. Gu, F. Xu, Valorization of food
640 waste impurities by catalytic co-pyrolysis for production of pyrolysis oil with high energy
641 potential, *J. Anal. Appl. Pyrolysis* 170 (2023) 105918. <https://doi.org/10.1016/j.jaap.2023.105918>

642 [19] H.V. Ly, B. Kwon, J. Kim, C. Oh, H.T. Hwang, J.S. Lee, S.-S. Kim, Effects of torrefaction on
643 product distribution and quality of bio-oil from food waste pyrolysis in N₂ and CO₂, *Waste*
644 *Manage. (Oxford)* 141 (2022) 16-26. <https://doi.org/10.1016/j.wasman.2022.01.013>

645 [20] T.J. Barzee, C. Yothers, A. Edalati, K. Rude, A. Chio, H.M. El Mashad, A. Franz, R. Zhang,
646 Pilot microalgae cultivation using food waste digestate with minimal resource inputs, *Bioresour.*
647 *Technol. Rep.* 19 (2022) 101200. <https://doi.org/10.1016/j.biteb.2022.101200>

648 [21] J. Li, L. Li, M. Suvarna, L. Pan, M. Tabatabaei, Y.S. Ok, X. Wang, Wet wastes to bioenergy
649 and biochar: A critical review with future perspectives, *Sci. Total Environ.* 817 (2022) 152921.
650 <https://doi.org/10.1016/j.scitotenv.2022.152921>

651 [22] C. Linyi, Q. Yujie, C. Buqing, W. Chenglong, Z. Shaohong, C. Renglu, Y. Shaohua, Y. Lan,
652 L. Zhiju, Enhancing degradation and biogas production during anaerobic digestion of food waste

653 using alkali pretreatment, Environ. Res. 188 (2020) 109743.
654 <https://doi.org/10.1016/j.envres.2020.109743>

655 [23] T.-H. Chen, M.-Y. Shen, C.-Y. Chen, Y.-W. Chen, L.-H. Wang, C.-Y. Chu, M.-C. Lee, H.-L.
656 Sun, Biogas production from food waste hydrolysate using a subcritical water pretreated process
657 and pulp wastewater seed sludge, Sustain. Energy Technol. Assess. 59 (2023) 103392.
658 <https://doi.org/10.1016/j.seta.2023.103392>

659 [24] W. Nurjuwita, A. Sasongko, T.J. Hartanto, M. Purwanto, Potential and characterization biogas
660 from tofu liquid waste with addition cow dung and effective microorganisms 4 as biocatalyst,
661 Materials Today: Proceedings 46 (2021) 1908-1912. <https://doi.org/10.1016/j.matpr.2021.02.025>

662 [25] K. Latha, R. Velraj, P. Shanmugam, S. Sivanesan, Mixing strategies of high solids anaerobic
663 co-digestion using food waste with sewage sludge for enhanced biogas production, J. Clean. Prod.
664 210 (2019) 388-400. <https://doi.org/10.1016/j.jclepro.2018.10.219>

665 [26] A. Tayyab, Z. Ahmad, T. Mahmood, A. Khalid, S. Qadeer, S. Mahmood, S. Andleeb, M.
666 Anjum, Anaerobic co-digestion of catering food waste utilizing Parthenium hysterophorus as co-
667 substrate for biogas production, Biomass Bioenergy 124 (2019) 74-82.
668 <https://doi.org/10.1016/j.biombioe.2019.03.013>

669 [27] T. Yuan, R. Sun, M. Shao, Q. Chen, Y. Lin, Q. Xu, Biochar regulates enzymes activity and
670 interspecies electron transfer to promote bioenergy recovery from a continuous two-stage food
671 waste anaerobic digestion process, J. Clean. Prod. 385 (2023) 135690.
672 <https://doi.org/10.1016/j.jclepro.2022.135690>

673 [28] T.-H. Kim, D. Song, Y.-J. Jeon, O. Hwang, J.-Y. Nam, Y.-M. Yun, Enhanced production of
674 biohydrogen and biomethane through a two-stage anaerobic fermentation of food waste mixed

675 with conductive additives, Chem. Eng. J. 476 (2023) 146520.
676 <https://doi.org/10.1016/j.cej.2023.146520>

677 [29] J.A. Lalman, S.R. Chaganti, C. Moon, D.-H. Kim, Elucidating acetogenic H₂ consumption in
678 dark fermentation using flux balance analysis, Bioresour. Technol. 146 (2013) 775-778.
679 <https://doi.org/10.1016/j.biortech.2013.07.125>

680 [30] Y.-M. Yun, M.-K. Lee, S.-W. Im, A. Marone, E. Trably, S.-R. Shin, M.-G. Kim, S.-K. Cho,
681 D.-H. Kim, Biohydrogen production from food waste: current status, limitations, and future
682 perspectives, Bioresour. Technol. 248 (2018) 79-87.
683 <https://doi.org/10.1016/j.biortech.2017.06.107>

684 [31] P. Zhou, E. Elbeshbishy, G. Nakhla, Optimization of biological hydrogen production for
685 anaerobic co-digestion of food waste and wastewater biosolids, Bioresour. Technol. 130 (2013)
686 710-718. <https://doi.org/10.1016/j.biortech.2012.12.069>

687 [32] J. Rajesh Banu, J. Merrylin, T.M. Mohamed Usman, R. Yukesh Kannah, M. Gunasekaran, S.-
688 H. Kim, G. Kumar, Impact of pretreatment on food waste for biohydrogen production: A review,
689 Int. J. Hydrogen Energy 45(36) (2020) 18211-18225.
690 <https://doi.org/10.1016/j.ijhydene.2019.09.176>

691 [33] D.-H. Kim, S.-H. Kim, H.-S. Shin, Hydrogen fermentation of food waste without inoculum
692 addition, Enzyme Microb. Technol. 45(3) (2009) 181-187.
693 <https://doi.org/10.1016/j.enzmictec.2009.06.013>

694 [34] S.H. Lee, S. Lee, S.-M. Lee, J. Cha, H.S. Lee, S.G. Kang, Biohydrogen Production from Food
695 Waste Using Glucose-Adapted Hyperthermophilic Archaeon, Waste Biomass Valor. 14(9) (2023)
696 2923-2930. <https://doi.org/10.1007/s12649-023-02049-z>

- 697 [35] G.K. Dinesh, R. Chauhan, S. Chakma, Influence and strategies for enhanced biohydrogen
698 production from food waste, *Renew. Sustain. Energy Rev.* 92 (2018) 807-822.
699 <https://doi.org/10.1016/j.rser.2018.05.009>
- 700 [36] S. Xue, H. Chen, F. Wang, G. Lv, L. Tan, G. Liu, The effect of substrate acidification on the
701 biohydrogen production by dark fermentation, *Int. J. Hydrogen Energy* 49 (2024) 177-188.
702 <https://doi.org/10.1016/j.ijhydene.2023.07.183>
- 703 [37] L. Regueira-Marcos, O. García-Depraect, R. Muñoz, Elucidating the role of pH and total
704 solids content in the co-production of biohydrogen and carboxylic acids from food waste via
705 lactate-driven dark fermentation, *Fuel* 338 (2023) 127238.
706 <https://doi.org/10.1016/j.fuel.2022.127238>
- 707 [38] J. Maroušek, O. Strunecký, V. Bartoš, M. Vochozka, Revisiting competitiveness of hydrogen
708 and algae biodiesel, *Fuel* 328 (2022) 125317. <https://doi.org/10.1016/j.fuel.2022.125317>
- 709 [39] H.M. Kadlimatti, B. Raj Mohan, M.B. Saidutta, Bio-oil from microwave assisted pyrolysis of
710 food waste-optimization using response surface methodology, *Biomass Bioenergy* 123 (2019) 25-
711 33. <https://doi.org/10.1016/j.biombioe.2019.01.014>
- 712 [40] A. Ben Hassen Trabelsi, K. Zaafour, W. Baghdadi, S. Naoui, A. Ouerghi, Second generation
713 biofuels production from waste cooking oil via pyrolysis process, *Renew. Energy* 126 (2018) 888-
714 896. <https://doi.org/10.1016/j.renene.2018.04.002>
- 715 [41] J. Zhao, Z. Wang, J. Li, B. Yan, G. Chen, Pyrolysis of food waste and food waste solid
716 digestate: A comparative investigation, *Bioresour. Technol.* 354 (2022) 127191.
717 <https://doi.org/10.1016/j.biortech.2022.127191>

- 718 [42] A. Raizada, S. Yadav, M. Tripathi, S. Misra, P. Mohanty, Food waste treatment using in situ
719 gasification after pyrolysis to produce hydrogen-rich syngas, *Biomass Convers. Bioref.* 13(11)
720 (2023) 9689-9699. <https://doi.org/10.1007/s13399-021-01857-4>
- 721 [43] B. Motavaf, P.E. Savage, Effect of process variables on food waste valorization via
722 hydrothermal liquefaction, *ACS ES&T Engineering* 1(3) (2021) 363-374.
723 <https://doi.org/10.1021/acsestengg.0c00115>
- 724 [44] M. Carmona-Cabello, I. García, A. Papadaki, E. Tsouko, A. Koutinas, M. Dorado, Biodiesel
725 production using microbial lipids derived from food waste discarded by catering services,
726 *Bioresour. Technol.* 323 (2021) 124597. <https://doi.org/10.1016/j.biortech.2020.124597>
- 727 [45] X. Ma, Z. Gao, M. Gao, Y. Ma, H. Ma, M. Zhang, Y. Liu, Q. Wang, Microbial lipid production
728 from food waste saccharified liquid and the effects of compositions, *Energy Convers. Manag.* 172
729 (2018) 306-315. <https://doi.org/10.1016/j.enconman.2018.07.005>
- 730 [46] F. Bibi, A. Jamal, Z. Huang, M. Urynowicz, M.I. Ali, Advancement and role of abiotic stresses
731 in microalgae biorefinery with a focus on lipid production, *Fuel* 316 (2022) 123192.
732 <https://doi.org/10.1016/j.fuel.2022.123192>
- 733 [47] R. Kakarla, J.W. Choi, J.H. Yun, B.H. Kim, J. Heo, S. Lee, D.H. Cho, R. Ramanan, H.S. Kim,
734 Application of high-salinity stress for enhancing the lipid productivity of *Chlorella sorokiniana*
735 HS1 in a two-phase process, *J. Microbiol.* 56(1) (2018) 56-64. [https://doi.org/10.1007/s12275-](https://doi.org/10.1007/s12275-018-7488-6)
736 [018-7488-6](https://doi.org/10.1007/s12275-018-7488-6)
- 737 [48] P. Swain, A. Tiwari, A. Pandey, Enhanced lipid production in *Tetraselmis* sp. by two stage
738 process optimization using simulated dairy wastewater as feedstock, *Biomass Bioenergy*
739 139(August 2019) (2020) 105643-105643. <https://doi.org/10.1016/j.biombioe.2020.105643>

740 [49] Y. Wang, S.H. Ho, C.L. Cheng, W.Q. Guo, D. Nagarajan, N.Q. Ren, D.J. Lee, J.S. Chang,
741 Perspectives on the feasibility of using microalgae for industrial wastewater treatment, *Bioresour.*
742 *Technol.* 222 (2016) 485-497. <https://doi.org/10.1016/j.biortech.2016.09.106>

743 [50] M. Mondal, S. Goswami, A. Ghosh, G. Oinam, O. Tiwari, P. Das, K. Gayen, M. Mandal, G.
744 Halder, Production of biodiesel from microalgae through biological carbon capture: a review, 3
745 *Biotech* 7 (2017) 1-21. <https://doi.org/10.1007/s13205-017-0727-4>

746 [51] D. Pleissner, W.C. Lam, Z. Sun, C.S.K. Lin, Food waste as nutrient source in heterotrophic
747 microalgae cultivation, *Bioresour. Technol.* 137 (2013) 139-146.
748 <https://doi.org/10.1016/j.biortech.2013.03.088>

749 [52] O. Haske-Cornelius, T. Vu, C. Schmiedhofer, R. Vielnascher, M. Dielacher, V. Sachs, M.
750 Grasmug, S. Kromus, G. Guebitz, Cultivation of heterotrophic algae on enzymatically hydrolyzed
751 municipal food waste, *Algal Research* 50 (2020) 101993.
752 <https://doi.org/10.1016/j.algal.2020.101993>

753 [53] X. Wang, M.-M. Zhang, Z. Sun, S.-F. Liu, Z.-H. Qin, J.-H. Mou, Z.-G. Zhou, C.S.K. Lin,
754 Sustainable lipid and lutein production from *Chlorella* mixotrophic fermentation by food waste
755 hydrolysate, *J. Hazard. Mater.* 400 (2020) 123258. <https://doi.org/10.1016/j.jhazmat.2020.123258>

756 [54] Q. Al Abdallah, B.T. Nixon, J.R. Fortwendel, The enzymatic conversion of major algal and
757 cyanobacterial carbohydrates to bioethanol, *Front. Energy Res.* 4 (2016) 36.
758 <https://doi.org/10.3389/fenrg.2016.00036>

759 [55] K.P. Kallarakkal, K. Muthukumar, A. Alagarsamy, A. Pugazhendhi, S. Naina Mohamed,
760 Enhancement of biobutanol production using mixotrophic culture of *Oscillatoria* sp. in cheese
761 whey water, *Fuel* 284(April 2020) (2021) 119008-119008.
762 <https://doi.org/10.1016/j.fuel.2020.119008>

763 [56] M.A. de Carvalho Silvello, G.A. Gasparotto, G.F. Ferreira, L.O. Santos, L.V. Fregolente, R.
764 Goldbeck, Nutrient Optimization Strategy to Increase the Carbohydrate Content of *Chlorella*
765 *vulgaris* and Evaluation of Hydrolysis and Fermentation Performance, *Bioenerg. Res.* 16(4) (2023)
766 2058-2067. <https://doi.org/10.1007/s12155-023-10660-0>

767 [57] S.O. Ajala, M.L. Alexander, Assessment of *Chlorella vulgaris*, *Scenedesmus obliquus*, and
768 *Oocystis minuta* for removal of sulfate, nitrate, and phosphate in wastewater, *Int. J. Energy*
769 *Environ. Eng.* 11(3) (2020) 311-326. <https://doi.org/10.1007/s40095-019-00333-0>

770 [58] E.J. Kim, S. Kim, H.G. Choi, S.J. Han, Co-production of biodiesel and bioethanol using
771 psychrophilic microalga *Chlamydomonas* sp. KNM0029C isolated from Arctic sea ice,
772 *Biotechnol. Biofuels* 13(1) (2020) 1-13. <https://doi.org/10.1186/s13068-020-1660-z>

773 [59] S.Y. Lee, I. Khoiroh, D.-V.N. Vo, P. Senthil Kumar, P.L. Show, Techniques of lipid extraction
774 from microalgae for biofuel production: a review, *Environ. Chem. Lett.* 19 (2021) 231-251.
775 <https://doi.org/10.1007/s10311-020-01088-5>

776 [60] K. Karimi, M. Saidi, P. Moradi, A. Taheri Najafabadi, Biodiesel production from
777 *Nannochloropsis* microalgal biomass-derived oil: An experimental and theoretical study using the
778 RSM-CCD approach, *Can. J. Chem. Eng.* 101(10) (2023) 5600-5610.
779 <https://doi.org/10.1002/cjce.24863>

780 [61] N.H.M. Yasin, N.N.C. Aziz, M.B.A. Azmai, M.F.M. Hanapi, Transesterification method of
781 microalgae biomass to produce fatty acid methyl esters, *J. Chem. Technol. Biotechnol.* n/a(n/a)
782 (2023). <https://doi.org/10.1002/jctb.7338>

783 [62] M.V. Rodionova, A.M. Bozieva, S.K. Zharmukhamedov, Y.K. Leong, J. Chi-Wei Lan, A.
784 Veziroglu, T.N. Veziroglu, T. Tomo, J.-S. Chang, S.I. Allakhverdiev, A comprehensive review on

785 lignocellulosic biomass biorefinery for sustainable biofuel production, *Int. J. Hydrogen Energy*
786 47(3) (2022) 1481-1498. <https://doi.org/10.1016/j.ijhydene.2021.10.122>

787 [63] N.K.N. Al-Shorgani, H. Shukor, P. Abdeshahian, M.S. Kalil, W.M.W. Yusoff, A.A. Hamid,
788 Enhanced butanol production by optimization of medium parameters using *Clostridium*
789 *acetobutylicum* YM1, *Saudi J. Biol. Sci.* 25(7) (2018) 1308-1321.
790 <https://doi.org/10.1016/j.sjbs.2016.02.017>

791 [64] B.E. Condor, M.D.G. de Luna, Y.-H. Chang, J.-H. Chen, Y.K. Leong, P.-T. Chen, C.-Y. Chen,
792 D.-J. Lee, J.-S. Chang, Bioethanol production from microalgae biomass at high-solids loadings,
793 *Bioresour. Technol.* 363 (2022) 128002. <https://doi.org/10.1016/j.biortech.2022.128002>

794 [65] Z. Yirgu, S. Leta, A. Hussien, M.M. Khan, Pretreatment and optimization of reducing sugar
795 extraction from indigenous microalgae grown on brewery wastewater for bioethanol production,
796 *Biomass Convers. Bioref.* 13(8) (2023) 6831-6845. <https://doi.org/10.1007/s13399-021-01779-1>

797 [66] L. Vargas-Estrada, A. Longoria, E. Arenas, J. Moreira, P.U. Okoye, Y. Bustos-Terrones, P.
798 Sebastian, A review on current trends in biogas production from microalgae biomass and
799 microalgae waste by anaerobic digestion and co-digestion, *BioEnergy Res.* 15(1) (2022) 77-92.
800 <https://doi.org/10.1007/s12155-021-10276-2>

801 [67] C. Xiao, Q. Liao, Q. Fu, Y. Huang, A. Xia, W. Shen, H. Chen, X. Zhu, Exergy analyses of
802 biogas production from microalgae biomass via anaerobic digestion, *Bioresour. Technol.* 289
803 (2019) 121709. <https://doi.org/10.1016/j.biortech.2019.121709>

804 [68] H.M. Zabed, S. Akter, J. Yun, G. Zhang, Y. Zhang, X. Qi, Biogas from microalgae:
805 Technologies, challenges and opportunities, *Renew. Sustain. Energy Rev.* 117 (2020) 109503.
806 <https://doi.org/10.1016/j.rser.2019.10950>

807 [69] N. Tasnim Sahrin, K. Shiong Khoo, J. Wei Lim, R. Shamsuddin, F. Musa Ardo, H. Rawindran,
808 M. Hassan, W. Kiatkittipong, E. Alaaeldin Abdelfattah, W. Da Oh, C. Kui Cheng, Current
809 perspectives, future challenges and key technologies of biohydrogen production for building a
810 carbon-neutral future: A review, *Bioresour. Technol.* 364 (2022) 128088.
811 <https://doi.org/10.1016/j.biortech.2022.128088>

812 [70] N. Fakhimi, M.J. Torres, E. Fernández, A. Galván, A. Dubini, D. González-Ballester,
813 *Chlamydomonas reinhardtii* and *Microbacterium forte* sp. nov., a mutualistic association that
814 favors sustainable hydrogen production, *Sci. Total Environ.* 913 (2024) 169559.
815 <https://doi.org/10.1016/j.scitotenv.2023.169559>

816 [71] P. Sharma, A. Jain, B.J. Bora, D. Balakrishnan, P.L. Show, R. Ramaraj, Ü. Ağbulut, K.S.
817 Khoo, Application of modern approaches to the synthesis of biohydrogen from organic waste, *Int.*
818 *J. Hydrogen Energy* 48(55) (2023) 21189-21213. <https://doi.org/10.1016/j.ijhydene.2023.03.029>

819 [72] D. Nagarajan, J.-s. Chang, D.-j. Lee, Pretreatment of microalgal biomass for efficient
820 biohydrogen production – Recent insights and future perspectives, *Bioresour. Technol.*
821 302(January) (2020) 122871-122871. <https://doi.org/10.1016/j.biortech.2020.122871>

822 [73] D. Kazawadi, J. Ntalikwa, G. Kombe, A. Saydut, A Review of Intermediate Pyrolysis as a
823 Technology of Biomass Conversion for Coproduction of Biooil and Adsorption Biochar, *J. Renew.*
824 *Energy* 2021 (2021) 1-10. <https://doi.org/10.1155/2021/5533780>

825 [74] X. Gong, B. Zhang, Y. Zhang, Y. Huang, M. Xu, Investigation on pyrolysis of low lipid
826 microalgae *Chlorella vulgaris* and *Dunaliella salina*, *Energy & fuels* 28(1) (2014) 95-103.
827 <https://doi.org/10.1021/ef401500z>

828 [75] X.J. Lee, H.C. Ong, Y.Y. Gan, W.-H. Chen, T.M.I. Mahlia, State of art review on conventional
829 and advanced pyrolysis of macroalgae and microalgae for biochar, bio-oil and bio-syngas

830 production, Energy Convers. Manag. 210 (2020) 112707.
831 <https://doi.org/10.1016/j.enconman.2020.112707>

832 [76] A.F. Ferreira, A.P. Soares Dias, Pyrolysis of microalgae biomass over carbonate catalysts, J.
833 Chem. Technol. Biotechnol. 95(12) (2020) 3270-3279. <https://doi.org/10.1002/jctb.6506>

834 [77] Z. Huiru, Y. Yunjun, F. Liberti, B. Pietro, F. Fantozzi, Technical and economic feasibility
835 analysis of an anaerobic digestion plant fed with canteen food waste, Energy Convers. Manage.
836 180 (2019) 938-948. <https://doi.org/10.1016/j.enconman.2018.11.045>

837 [78] Y. Chen, L. Pinegar, J. Immonen, K.M. Powell, Conversion of food waste to renewable
838 energy: A techno-economic and environmental assessment, Journal of Cleaner Production 385
839 (2023) 135741. <https://doi.org/10.1016/j.jclepro.2022.135741>

840 [79] M. Melikoglu, A. Tekin, Biohydrogen production from food and agricultural wastes: A global
841 review and a techno-economic evaluation for Turkey, Int. J. Hydrogen Energy 62 (2024) 913-924.
842 <https://doi.org/10.1016/j.ijhydene.2024.03.173>

843 [80] M. Alherbawi, P. Parthasarathy, S. Elkhalfifa, T. Al-Ansari, G. McKay, Techno-economic and
844 environmental analyses of the pyrolysis of food waste to produce bio-products, Heliyon 10(6)
845 (2024). <https://doi.org/10.1016/j.heliyon.2024.e27713>

846 [81] N. Rajendran, J. Han, Techno-economic analysis of food waste valorization for integrated
847 production of polyhydroxyalkanoates and biofuels, Bioresour. Technol. 348 (2022) 126796.
848 <https://doi.org/10.1016/j.biortech.2022.126796>

849 [82] L. Amer, B. Adhikari, J. Pellegrino, Technoeconomic analysis of five microalgae-to-biofuels
850 processes of varying complexity, Bioresour. Technol. 102(20) (2011) 9350-9359.
851 <https://doi.org/10.1016/j.biortech.2011.08.010>

852 [83] S. Dutta, F. Neto, M.C. Coelho, Microalgae biofuels: A comparative study on techno-
853 economic analysis & life-cycle assessment, *Algal research* 20 (2016) 44-52.
854 <https://doi.org/10.1016/j.algal.2016.09.018>

855 [84] I. Pikaar, S. Matassa, B.L. Bodirsky, I. Weindl, F. Humpenöder, K. Rabaey, N. Boon, M.
856 Bruschi, Z. Yuan, H. van Zanten, Decoupling livestock from land use through industrial feed
857 production pathways, *Environ. Sci. Technol.* 52(13) (2018) 7351-7359.
858 <https://doi.org/10.1021/acs.est.8b00216>

859 [85] S. FUNG, Sustainable development goals, Available at this link: [https://www.un.](https://www.un.org/sustainabledevelopment/inequality)
860 [org/sustainabledevelopment/inequality](https://www.un.org/sustainabledevelopment/inequality) (2015).

861 [86] B. Baldassarre, J. Konietzko, P. Brown, G. Calabretta, N. Bocken, I.O. Karpen, E.J. Hultink,
862 Addressing the design-implementation gap of sustainable business models by prototyping: A tool
863 for planning and executing small-scale pilots, *J. Clean. Prod.* 255 (2020) 120295.
864 <https://doi.org/10.1016/j.jclepro.2020.120295>

865 [87] B. Hankamer, L. Pregelj, S. O’Kane, K. Hussey, D. Hine, Delivering impactful solutions for
866 the bioeconomy, *Trends Plant Sci.* 28(5) (2023) 583-596.
867 <https://doi.org/10.1016/j.tplants.2023.02.007>

868 [88] J. Fu, Y. Huang, Q. Liao, A. Xia, Q. Fu, X. Zhu, Photo-bioreactor design for microalgae: a
869 review from the aspect of CO₂ transfer and conversion, *Bioresour. Technol.* 292 (2019) 121947.
870 <https://doi.org/10.1016/j.biortech.2019.121947>

871 [89] O. García-Depraect, I. Mirzazada, L.J. Martínez-Mendoza, L. Regueira-Marcos, R. Muñoz,
872 Biotic and abiotic insights into the storage of food waste and its effect on biohydrogen and methane
873 production potential, *J. Water Process Eng.* 53 (2023) 103840.
874 <https://doi.org/10.1016/j.jwpe.2023.103840>

- 875 [90] T. Hai, P. Mishra, J.M. Zain, K. Saini, N.M. Kumar, Z. Ab Wahid, Co-digestion of domestic
876 kitchen food waste and palm oil mill effluent for biohydrogen production, *Sustain. Energy*
877 *Technol. Assess.* 55 (2023) 102965. <https://doi.org/10.1016/j.seta.2022.102965>
- 878 [91] S. Neha, N. Remya, Co-production of biooil and biochar from microwave co-pyrolysis of
879 food-waste and plastic using recycled biochar as microwave susceptor, *Sustain. Energy Technol.*
880 *Assess.* 54 (2022) 102892. <https://doi.org/10.1016/j.seta.2022.102892>
- 881 [92] M. Chen, S. Zhang, Y. Su, X. Niu, S. Zhu, X. Liu, Catalytic co-pyrolysis of food waste
882 digestate and corn husk with CaO catalyst for upgrading bio-oil, *Renew. Energy* 186 (2022) 105-
883 114. <https://doi.org/10.1016/j.renene.2021.12.139>
- 884 [93] S. Neha, N. Remya, Optimization of bio-oil production from microwave co-pyrolysis of food
885 waste and low-density polyethylene with response surface methodology, *J. Environ. Manage.* 297
886 (2021) 113345. <https://doi.org/10.1016/j.jenvman.2021.113345>
- 887 [94] J.F. García-Martín, C.C. Barrios, F.-J. Alés-Álvarez, A. Dominguez-Sáez, P. Alvarez-Mateos,
888 Biodiesel production from waste cooking oil in an oscillatory flow reactor. Performance as a fuel
889 on a TDI diesel engine, *Renewable Energy* 125 (2018) 546-556.
890 <https://doi.org/10.1016/j.renene.2018.03.002>
- 891 [95] J. Umamaheswari, M.S. Kavitha, S. Shanthakumar, Outdoor cultivation of *Chlorella*
892 *pyrenoidosa* in paddy-soaked wastewater and a feasibility study on biodiesel production from wet
893 algal biomass through in-situ transesterification, *Biomass Bioenergy* 143 (2020) 105853.
894 <https://doi.org/10.1016/j.biombioe.2020.105853>
- 895 [96] T.T. Mamo, Y.S. Mekonnen, Microwave-Assisted Biodiesel Production from Microalgae,
896 *Scenedesmus* Species, Using Goat Bone-Made Nano-catalyst, *Appl. Biochem. Biotechnol.* 190(4)
897 (2020) 1147-1162. <https://doi.org/10.1007/s12010-019-03149-0>

898 [97] S.S. de Jesus, G.F. Ferreira, L.S. Moreira, R.M. Filho, Biodiesel production from microalgae
899 by direct transesterification using green solvents, *Renewable Energy* 160 (2020) 1283-1294.
900 <https://doi.org/10.1016/j.renene.2020.07.056>

901 [98] P.I.G. Acebu, M.D.G. de Luna, C.-Y. Chen, R.R.M. Abarca, J.-H. Chen, J.-S. Chang,
902 Bioethanol production from *Chlorella vulgaris* ESP-31 grown in unsterilized swine wastewater,
903 *Bioresour. Technol.* 352 (2022) 127086. <https://doi.org/10.1016/j.biortech.2022.127086>

904 [99] G.M. Figueroa-Torres, W.M.A. Wan Mahmood, J.K. Pittman, C. Theodoropoulos, Microalgal
905 biomass as a biorefinery platform for biobutanol and biodiesel production, *Biochem. Eng. J.* 153
906 (2020) 107396-107396. <https://doi.org/10.1016/j.bej.2019.107396>

907 [100] T.Y. Tsai, Y.C. Lo, C.D. Dong, D. Nagarajan, J.S. Chang, D.J. Lee, Biobutanol production
908 from lignocellulosic biomass using immobilized *Clostridium acetobutylicum*, *Appl. Energy*
909 277(July) (2020) 115531-115531. <https://doi.org/10.1016/j.apenergy.2020.115531>

910 [101] G. Narchonai, C. Arutselvan, F. LewisOscar, N. Thajuddin, Enhancing starch
911 accumulation/production in *Chlorococcum humicola* through sulphur limitation and 2,4- D
912 treatment for butanol production, *Biotechnol. Rep.* 28 (2020) e00528-e00528.
913 <https://doi.org/10.1016/j.btre.2020.e00528>

914 [102] H. Wu, J. Li, Q. Liao, Q. Fu, Z. Liu, Enhanced biohydrogen and biomethane production
915 from *Chlorella* sp . with hydrothermal treatment, *Energy Convers. Manag.* 205(December 2019)
916 (2020) 112373-112373. <https://doi.org/10.1016/j.enconman.2019.112373>

917 [103] A. Jehlee, S. Rodjaroen, J. Waewsak, A. Reungsang, S. O-Thong, Improvement of
918 biohythane production from *Chlorella* sp. TISTR 8411 biomass by co-digestion with organic
919 wastes in a two-stage fermentation, *Int. J. Hydrogen Energy* 44(32) (2019) 17238-17247.
920 <https://doi.org/10.1016/j.ijhydene.2019.03.026>

921 [104] P. Ayala-Parra, Y. Liu, J.A. Field, R. Sierra-Alvarez, Nutrient recovery and biogas generation
922 from the anaerobic digestion of waste biomass from algal biofuel production, *Renew. Energy* 108
923 (2017) 410-416. <https://doi.org/10.1016/j.renene.2017.02.085>

924 [105] V. Kinnunen, J. Rintala, The effect of low-temperature pretreatment on the solubilization
925 and biomethane potential of microalgae biomass grown in synthetic and wastewater media,
926 *Bioresour. Technol.* 221 (2016) 78-84. <https://doi.org/10.1016/j.biortech.2016.09.017>

927 [106] G. Zhen, X. Lu, T. Kobayashi, G. Kumar, K. Xu, Anaerobic co-digestion on improving
928 methane production from mixed microalgae (*Scenedesmus* sp., *Chlorella* sp.) and food waste:
929 Kinetic modeling and synergistic impact evaluation, *Chem. Eng. J.* 299 (2016) 332-341.
930 <https://doi.org/10.1016/j.cej.2016.04.118>

931 [107] S. Satheesh, A. Pugazhendi, B.A. Al-Mur, R. Balasubramani, Biohydrogen production
932 coupled with wastewater treatment using selected microalgae, *Chemosphere* 334 (2023) 138932.
933 <https://doi.org/10.1016/j.chemosphere.2023.138932>

934 [108] P. Sivagurunathan, G. Kumar, T. Kobayashi, K. Xu, S.H. Kim, D.D. Nguyen, S.W. Chang,
935 Co-digestion of untreated macro and microalgal biomass for biohydrogen production: Impact of
936 inoculum augmentation and microbial insights, *Int. J. Hydrogen Energy* 43(25) (2018) 11484-
937 11492. <https://doi.org/10.1016/j.ijhydene.2018.02.193>

938 [109] G. Kumar, P. Sivagurunathan, N.B.D. Thi, G. Zhen, T. Kobayashi, S.H. Kim, K. Xu,
939 Evaluation of different pretreatments on organic matter solubilization and hydrogen fermentation
940 of mixed microalgae consortia, *Int. J. Hydrogen Energy* 41(46) (2016) 21628-21640.
941 <https://doi.org/10.1016/j.ijhydene.2016.05.195>

942 [110] A.K. Sharma, P. Ghodke, P.K. Sharma, S. Manna, A. Pugazhendhi, L. Matsakas, A. Patel,
943 Holistic utilization of *Chlorella pyrenoidosa* microalgae for extraction of renewable fuels and

944 value-added biochar through in situ transesterification and pyrolysis reaction process, *Biomass*
945 *Convers. Bioref.* 14(4) (2024) 5261-5274. <https://doi.org/10.1007/s13399-022-02713-9>
946 [111] T. Aysu, O. Ola, M.M. Maroto-Valer, A. Sanna, Effects of titania based catalysts on in-situ
947 pyrolysis of Pavlova microalgae, *Fuel Process. Technol.* 166 (2017) 291-298.
948 <https://doi.org/10.1016/j.fuproc.2017.05.001>

949

950 **Table Titles**

951 **Table 1.** Recent studies on the direct conversion of food waste into biofuels.

952 **Table 2.** Recent studies on the conversion methods of microalgal biomass into different forms of
953 bioenergy. These include biodiesel (transesterification), alcohol-based biofuel (fermentation),
954 biomethane (anaerobic digestion), biohydrogen (fermentation), bio-oil, biochar and bio-syngas
955 (pyrolysis).

956

957

958 **Figure Captions**

959 **Figure 1.** Direct conversion approaches of food waste to biofuels.

960 **Figure 2.** Indirect transformation methods through the cultivation of microorganism grown on
961 food waste for the generation of biofuels.

962 **Figure 3.** Photosynthesis process of microalgae cell assimilating light, water, and organic
963 substrates from food waste for the synthesis of intracellular products. Adapted and modified from
964 [88].

965 **Figure 4.** Biorefinery of microalgae for biofuel production using different processes. The
966 techniques include esterification/transesterification, fermentation, anaerobic digestion,
967 fermentation, and pyrolysis.

968 **Figure 5.** Historical events and prediction for microalgae cultivation since its initiation in 2000 –
969 2050 for the production of biofuel product and the correlation with SDGs.

970 **Table 1.** Recent studies on the direct conversion of food waste into biofuels.

| Technology | Food waste (co-substrate) | Food waste pretreatment | Inoculum | Process conditions | Biofuel | Production level | Reference |
|---------------------|---|---|---|---|---------|---|-----------|
| Anaerobic digestion | Restaurant food waste | Drying, milling, subcritical water hydrolysis | Seed sludge from a pulp wastewater treatment plant | Batch, 37 °C, 120 rpm | Biogas | 807 mL g ⁻¹ VS (329.5 mL CH ₄ g ⁻¹ VS) | [23] |
| Anaerobic digestion | Food waste prepared by mixing 20% meat, 15% tofu, 30% vegetables, 20% rice, 15% steamed bread | Blending | Seed sludge from an anaerobic digester | Continuous two-stage anaerobic digestion, 37 °C, addition of biochar | Biogas | Specific energy yields up to 1.23 kJ g ⁻¹ VS d ⁻¹ g ⁻¹ biochar | [27] |
| Anaerobic digestion | Cafeteria food waste | Blending, heat shock at 90 °C for 20 min, cooling at 4 °C for 1 h | Seed sludge from a brewery wastewater treatment plant | Batch, 38 °C, pH 5.5, 270 rpm, absence of light, addition of PAC catalyst | Biogas | 277 mL CH ₄ g ⁻¹ COD (1446 mL in total) | [28] |
| Anaerobic digestion | Canteen food waste | Blending, alkaline treatment using 1 % CaO | Inoculum from a municipal wastewater treatment plant | Fed-batch, 37 °C, 30 d solid retention time | Biogas | 829 mL g ⁻¹ VS (65.5% CH ₄) | [22] |
| Anaerobic digestion | Catering food waste (<i>Parthenium hysterophorus</i>) | Grinding (drying, microwave irradiation, steam sterilization) | Cattle manure | Batch, 30 °C | Biogas | 559 ml L ⁻¹ d ⁻¹ (5532 ml L ⁻¹ in total) | [26] |

| | | | | | | | |
|------------------------|---|---|--|---|------------------------------|---|------|
| Anaerobic fermentation | Food waste from food waste treatment plant | for co-substrate) Sorting, shredding | Inoculum from a lake in a China park | 37 °C | Hydrogen | 120.78 mL g ⁻¹ VS | [36] |
| Anaerobic fermentation | Food waste prepared by mixing 78 % potato, 14 % chicken, 4 % lard, 4 % cabbage | Grinding, storage at -20 °C | Digestate from an anaerobic digester | 35 °C | Hydrogen | 9.7 NL (L d) ⁻¹ | [89] |
| Anaerobic fermentation | Food waste prepared by mixing 78% potato flakes, 14% chicken breast, 4% cabbage, 4% pork lard | Storage at -20 °C | Digestate from an anaerobic digester | 37 °C, 200 rpm | Hydrogen | 103.4 NmL g ⁻¹ VS | [37] |
| Anaerobic fermentation | Potato peel waste from a local restaurant | Drying, grinding, freeze storage | <i>T. onnurineus</i> NA1 | 80 °C, 60 min, pH 6.1 – 6.2 | Hydrogen | 1.29 mmol L ⁻¹ h ⁻¹ | [34] |
| Anaerobic fermentation | Kitchen food waste (palm oil mill effluent (POME)) | Blending, sterilization | Immobilized <i>Bacillus anthracis</i> PUNAJAN1 | 35 °C, pH 5.0 | Hydrogen | 67 mL h ⁻¹ | [90] |
| Pyrolysis | Food waste prepared by mixing 30% vegetables, 70% cereals (trash plastic bag) | Not stated | - ^a | Microwave at 800W, 550 °C, 7 s | Bio-oil, biochar, bio-syngas | 40 – 52% bio-oil, 30 – 46% biochar, 10 – 30% bio-syngas | [91] |
| Pyrolysis | Food waste digestate (corn husk) | Drying, grinding | - ^a | 600 °C, ratio of food waste digestate to corn husk at 4:1 | Bio-oil, biochar, bio-syngas | 15.25% bio-oil, 52.75% biochar, 30.5% bio-syngas | [92] |

| | | | | | | | |
|---------------------|---|--|----------------|---|------------------------------|--|------|
| Pyrolysis | Household food waste (13% trash plastic bag) | Drying, grinding | - ^a | (w/w), addition of CaO catalyst Microwave at 1000 W, 550 °C, 7 s, addition of 6 wt% of granular activated carbon | Bio-oil, biochar, bio-syngas | 42% bio-oil, 42% biochar, 16% bio-syngas | [93] |
| Pyrolysis | Food waste from hostels and hotels | Drying, pulverization, sieving | - ^a | Microwave at 450W, 400 °C, 30 min, 50 mL min ⁻¹ N ₂ purging | Bio-oil | 30.2% bio-oil | [39] |
| Pyrolysis | Waste cooking oil | None | - ^a | 800 °C with a heating rate of 15 °C min ⁻¹ | Bio-oil, biochar | 80% bio-oil, 20% biochar | [40] |
| Transesterification | Lipid from bakery waste | Fungal hydrolysis, heating for water removal | - ^a | 60 °C, 2h, addition of CH ₃ OH and KOH | Biodiesel | 100% yield | [14] |
| Transesterification | Used mixture of olive oil and sunflower oil (1:1) from catering service | Filtration, heating for water removal | - ^a | 60 °C, 30 min., 0.67 Hz oscillatory frequency, addition of CH ₃ OH and NaOH | Biodiesel | 72% yield | [94] |

971 -^a indicates not applicable.

972 **Table 2.** Recent studies on the conversion methods of microalgal biomass into different forms of bioenergy. These include biodiesel
 973 (transesterification), alcohol-based biofuel (fermentation), biomethane (anaerobic digestion), biohydrogen (fermentation), bio-oil,
 974 biochar and bio-syngas (pyrolysis).

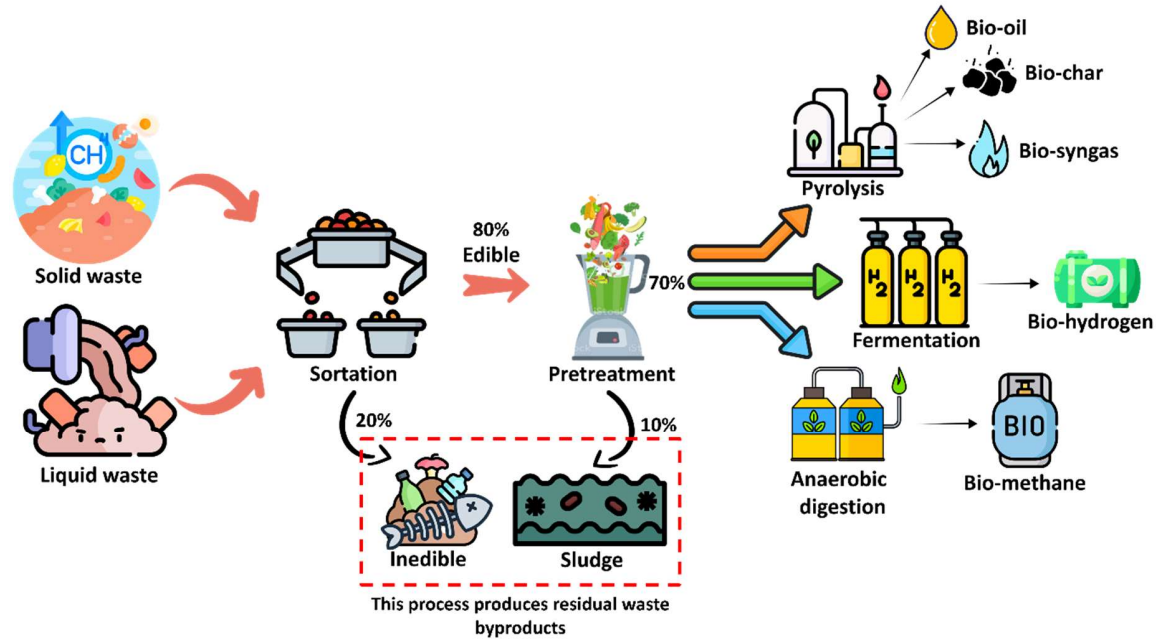
| Microalgae species | Conversion method | Process conditions | Biofuel | Conversion yield | Reference |
|--|---|--|------------|--|-----------|
| <i>Nannochloropsis</i> | <i>In-situ</i> transesterification using H ₂ SO ₄ (0.13wt% in methanol) | 69 °C, 30 min, ratio of methanol to biomass of 15:1 (v/w) | Biodiesel | 99% | [60] |
| <i>C. vulgaris</i> | <i>In-situ</i> transesterification using H ₂ SO ₄ | 85 °C, 2 h, 860 rpm, 12 mL methanol, 0.02 g dry biomass | Biodiesel | 243.48 mg FAME g ⁻¹ biomass | [61] |
| <i>C. pyrenoidosa</i> | <i>In-situ</i> transesterification using H ₂ SO ₄ (5% in methanol) | 90 °C, 60 min, ratio of hexane to wet biomass of 2:1 (v/w) | Biodiesel | 46.5% | [95] |
| <i>Scenedesmus</i> | Transesterification using CaO nano-catalyst prepared from goat bone after microwave-assisted lipid extraction | 60 °C, 3 h, ratio of methanol to microalgal oil of 11:1 | Biodiesel | 92% | [96] |
| <i>C. pyrenoidosa</i> | <i>In-situ</i> transesterification using H ₂ SO ₄ | 40 – 80 °C, 150 min, methyl ether as solvent | Biodiesel | 71 – 92% | [97] |
| <i>C. vulgaris</i> sp. ESP-31 | Fermentation using <i>S. cerevisiae</i> FAY-1 after acid hydrolysis pretreatment | Batch, 30 °C, 8 h, 200 rpm, 10 g L ⁻¹ biomass | Bioethanol | 84.2% | [98] |
| <i>Chlamydomonas reinhardtii</i> CCAP 11/32C | Fermentation using <i>C. acetobutylicum</i> DSM 792 after autoclave pretreatment | Batch, 37 °C, 20 h, pH 6.5, 10 g L ⁻¹ biomass | Biobutanol | 0.10 g butanol g ⁻¹ DCW | [99] |
| <i>Synechococcus elongates</i> PCC7942 (pigment extracted residue) | Fermentation using <i>C. acetobutylicum</i> | Batch, 37 °C, 30 – 40 h, 180 g L ⁻¹ biomass | Biobutanol | 0.09 g butanol g ⁻¹ biomass | [100] |

| | | | | | |
|---|--|---|---------------------------------|--|-------|
| <i>Chlorococcum humicola</i> | Fermentation using <i>C. acetobutylicum</i> 11274 after acid hydrolysis pretreatment | Batch, 30 °C, 72 h, pH 6.5 | Biobutanol | - ^a | [101] |
| <i>Chlorella</i> sp. | Two-stage anaerobic fermentation | 1 st stage: 37 °C, pH 6 2 nd stage: pH 7 | Biomethane, biohydrogen | 434 mL CH ₄ g ⁻¹ VS, 5.15 mL H ₂ g ⁻¹ VS | [102] |
| <i>Chlorella</i> sp. TISTR 8411 | Two-stage anaerobic fermentation | 1 st stage: 55 °C, 7 d 2 nd stage: 55 °C, 30 d | Biomethane, biohydrogen | 164 – 177 mL g ⁻¹ VS, 23 – 35 mL H ₂ g ⁻¹ VS | [103] |
| <i>C. sorokiniana</i> 1412 (lipid-extracted biomass) | Anaerobic digestion | 30 °C, 42 d | Biomethane | 220 – 280 mL CH ₄ g ⁻¹ VS | [104] |
| <i>C. vulgaris</i> and mixed culture of native algae species dominating by <i>Scenedesmus</i> sp. | Anaerobic digestion | 35 °C, 46 d | Biomethane | 154 – 252 mL CH ₄ g ⁻¹ VS | [105] |
| <i>Chlorella</i> sp. and <i>Scenedesmus</i> sp. (food waste) | Anaerobic digestion | 35 °C, 40 d | Biomethane | 640 mL CH ₄ g ⁻¹ VS | [106] |
| <i>C. pyrenoidosa</i> , <i>S. obliquus</i> , <i>C. sorokiniana</i> | Dark fermentation | 4 d | Biohydrogen | 45.5 mL H ₂ g ⁻¹ VS (<i>C. pyrenoidosa</i>) 38.4 mL H ₂ g ⁻¹ VS (<i>S. obliquus</i>) 34.8 mL H ₂ g ⁻¹ VS (<i>C. sorokiniana</i>) | [107] |
| Mixed microalgal biomass <i>Gelidium amansii</i> | Co-digestion | 35 °C, 6 d, pH 7.0 | Biohydrogen | 45 mL H ₂ g ⁻¹ DCW | [108] |
| Mixed microalgae consortia | Fermentation | 37 °C, 835 h | Biohydrogen | 38 mL H ₂ g ⁻¹ VS | [109] |
| <i>C. pyrenoidosa</i> | <i>In-situ</i> pyrolysis | 500 °C | Bio-oil, biochar, pyrolysis gas | 35.33wt% bio-oil, 51.23wt% biochar, | [110] |

| | | | | | | |
|-----|---|--------------------------|--|---------|---------------------------------------|-------|
| 975 | <i>Pavlova</i> sp. (catalyst Ni/TiO ₂) | <i>In-situ</i> pyrolysis | 500 °C, 60 min, 545 mL min ⁻¹ N ₂ purging | Bio-oil | 13.44wt% pyrolysis gas 22.55wt% | [111] |
|-----|---|--------------------------|--|---------|---------------------------------------|-------|

^a - indicates not available

976 **Figure 1**



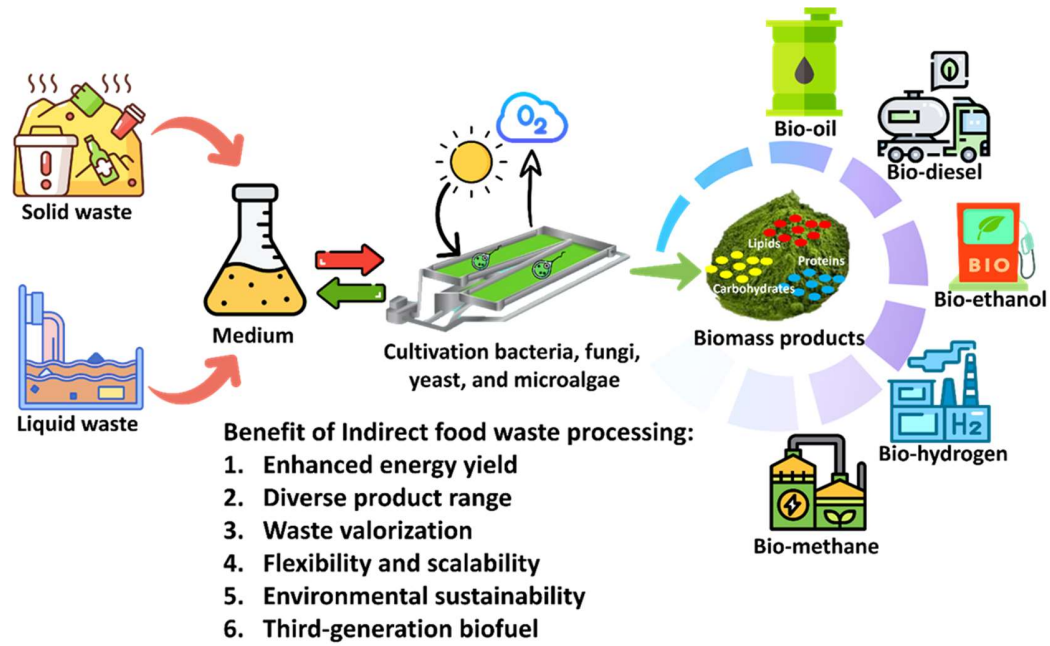
Benefit of direct food waste processing:

1. Efficiency resource utilization
2. Renewable energy generation
3. Waste reduction
4. Nutrient recovery
5. Economic benefit
6. Second-generation biofuel

977

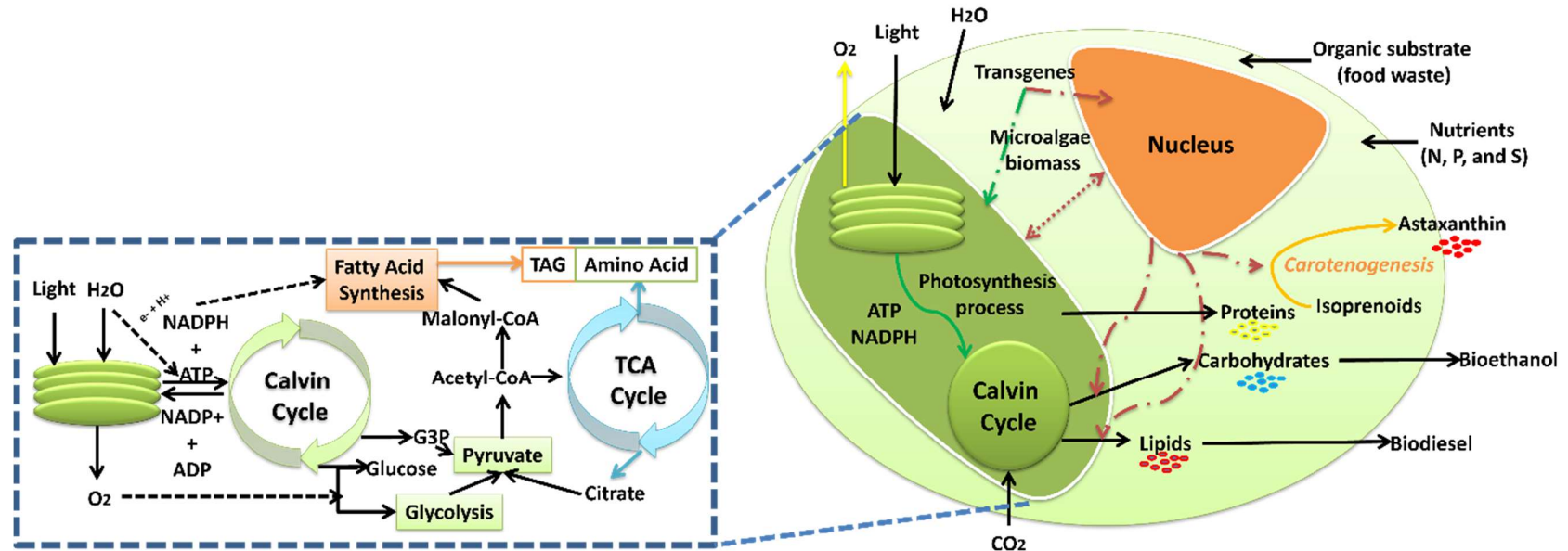
978

979 **Figure 2**

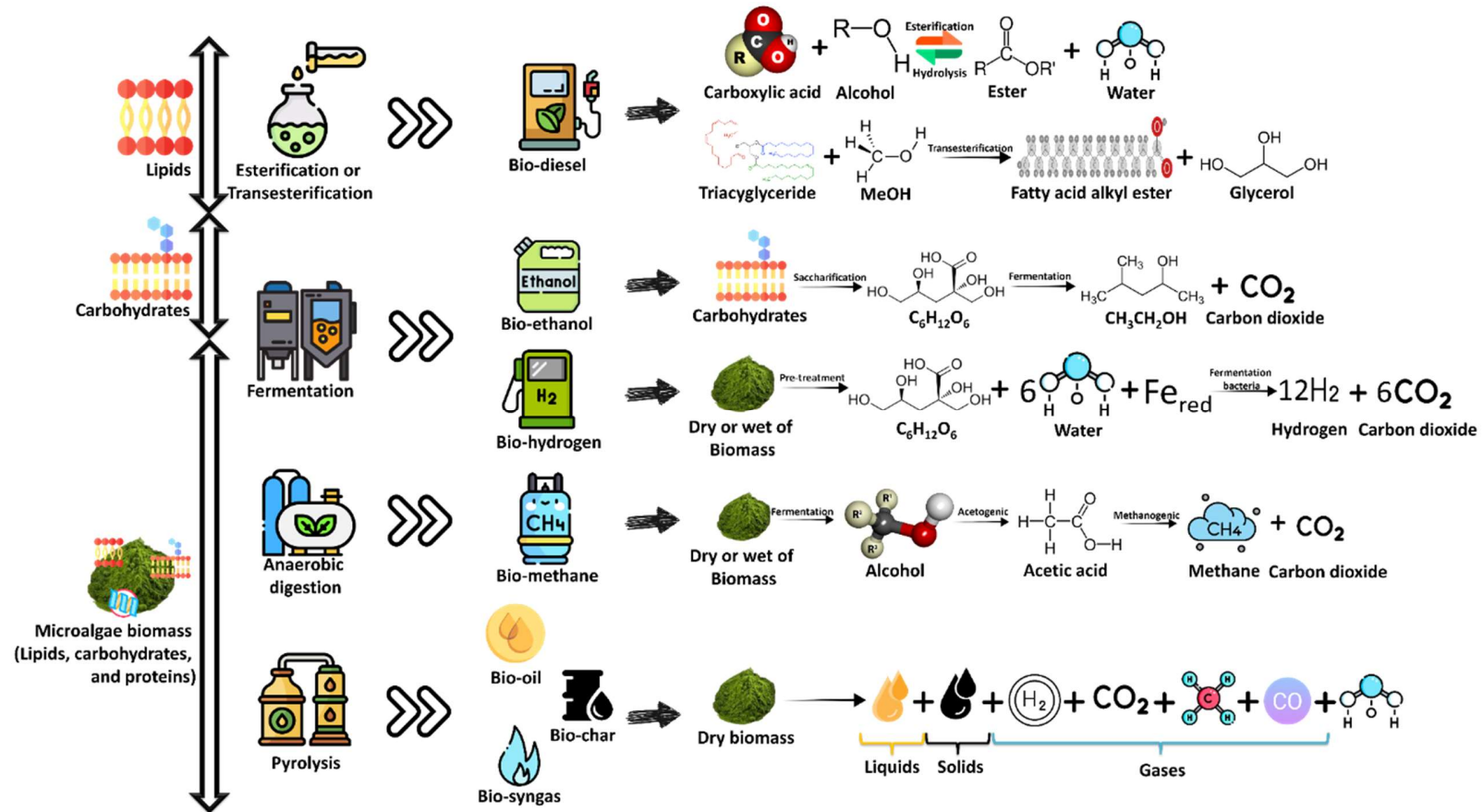


980

981 **Figure 3**



982



985 **Figure 5**

